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Advances in lung cancer basic and translational research in 2025 – Overview and perspectives focusing on non-small cell lung cancer

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Abstract

Basic and translational research in lung cancer is a rapidly evolving field with transformational impact in early detection, diagnosis, therapeutic development and personalization of care. Recent advances have greatly increased our understanding in the molecular genomics, proteomics, pathogenesis and cellular biology of this deadly malignancy. The International Association for the Study of Lung Cancer (IASLC) recently formed a Basic and Translational Science (BaTS) Committee to further enhance the scientific leadership of IASLC in thoracic cancer

research. This review by members of the committee highlights the breadth of current research in NSCLC, with a focus on molecular risk factors and processes in tumorigenesis, heterogeneity, phenotypic plasticity, metabolic reprogramming, immunobiology, the immune microenvironment and microbiome. This review also identifies future research areas that may lead to further improvement in survival outcomes and curative therapies especially for patients with advanced NSCLC.

Keywords

NSCLC; translational research; genomics; early lung cancer; states of art; tumor heterogeneity; cancer plasticity; immune microenvironment; TME

Introduction

Lung cancer is marked by complex genomic and epigenomic aberrations dysregulating the cellular signaling mechanisms and functions of tumor cells, and their interaction with the tumor microenvironment (TME). Non-small cell lung cancer (NSCLC) is commonly driven by a complex interplay of many biological pathways, including among others, signal transduction, DNA repair, chromatin modifications, metabolism, and immune checkpoint pathways. Breakthroughs that have occurred in basic and translational research during the last decades have not only enhanced our understanding of the biology and disease mechanisms in lung cancer, but they have also translated into the development of novel and personalized therapeutic strategies, which have improved the overall survival and quality of life for lung cancer patients. In this context, the Basic and Translational Science (BaTS) Committee of the International Association for the Study of Lung Cancer (IASLC) provides a review of the recent advances in our understanding of NSCLC at a cellular and molecular level, and its interactions with the tumor microenvironment (TME). We highlight the breadth of current lung cancer research and identify future directions towards advancing scientific innovation to improve patient outcomes and curative therapies.

Inherited susceptibility to lung cancer

Major progress has been made to elucidate both single-gene and polygenic inheritance of lung cancer.^{1,2} A significant proportion (4–15%) of patients with lung cancers harbor pathogenic germline variants in cancer genes.^{3,4,5} These are low-frequency variants with high or moderate-penetrance genes, such as *BRCA1*, *BRCA2*, *CHEK2*, *ATM*, *BAP1*, *EGFR* and *TP53*. The genome-wide association studies (GWAS) have identified more than 60 lung cancer susceptibility loci.^{1,6,7} The potential clinical applications of these GWAS results are the development of polygenic risk scores (PRSs), which aggregate information from multiple genetic variants to quantify an individual's genetic predisposition to lung cancer. Different PRSs were demonstrated as predictors of lung cancer independent of conventional clinical risk factors.^{8,9} A genome-wide PRS, leveraging the full GWAS data to quantify genetic predisposition, was recently demonstrated to outperform previously reported PRSs.¹⁰ Although the predictive power of the best-performing PRS does not surpass smoking history, it is comparable to other factors used in lung cancer risk models,

such as age and sex.^{8,10} Moreover, the polygenic background has been shown to modulate the risk associated with smoking. However, it remains controversial whether the risk stratification benefits of PRSs are more meaningful in never or ever-smokers.^{11,12,13,14} Nonetheless, advances in germline genetic testing and PRS provide new opportunities to identify individuals at high risk of lung cancer and we expect this emerging genetic knowledge to be increasingly integrated into lung cancer screening programs.^{15,16,17,2}

Carcinogenesis and early cancer interception

Early lung carcinogenesis is a complex process involving the sequential accumulation of molecular and genetic abnormalities. Putative pre-invasive lesions have been identified for NSCLC: squamous dysplasia and carcinoma in situ as precursors of lung squamous cell carcinomas (LUSC), and atypical adenomatous hyperplasia for lung adenocarcinomas (LUAD).¹⁸

Risk factors

The dominant risk factor associated with lung cancer is tobacco exposure, with over two thirds of cases attributable to smoking.¹⁹ With the rising incidence of lung cancer in non-smokers, additional risk factors have emerged including exposure to ionizing radiation (e.g. radon), occupational carcinogens (e.g. asbestos), and air pollution.²⁰ While mutagenic potential is a common feature amongst these exposures along with accumulating age-related mutations, chronic inflammation is likely also a critical component for lung cancer development and progression, as increased risk has been observed in chronic obstructive pulmonary disease and studies of particulate matter air pollution.^{21,22} At the same time, adaptive immune surveillance likely limits cancer development via recognition of mutations as foreign neo-antigens, evidenced for instance by increased lung cancer incidence in immune-suppressed populations and immunogenetic associations with cancer risk.^{23,24} As mentioned, family history is also an important risk factor. Taken together, these factors likely contribute in concert to influence lung cancer risk, but greater mechanistic understanding will provide further insights for improved cancer screening and prevention.

Squamous carcinogenesis

Lung squamous cell carcinoma (LUSC) accounts for 30–40% of lung cancers worldwide and its pathogenesis is significantly linked to tobacco smoking.²⁵ In North America, LUSC prevalence has dropped to ~20%, with more LUSC occurring in peripheral lung, in association with a decline in smoking.¹⁸ Central LUSCs are preceded by preneoplastic bronchial changes ranging from hyperplasia and metaplasia to mild, moderate, and severe dysplasia, culminating in carcinoma in situ (CIS) (Fig.1). Preinvasive lesions including dysplasia and CIS typically persist or progress locally within the bronchial tree and are often linked to the development of invasive cancer in distant lung areas, supporting the concept of field cancerization. Recent molecular studies have revealed significant chromosomal instability and methylation changes in bronchial preneoplastic lesions, leading to increased expression of cell cycle pathways and DNA damage response genes, along with transitory metabolic reprogramming and immune sensing and unleashing of tissue resident immune cells. In high-grade pre-invasive lesions, the activation and mobilization of immune cells

of both innate and adaptive immunity develop. However, a concomitant inhibition of immune response occurs before invasion²⁶ and is associated with the risk of progression.²⁷ Even in CIS lesions that regress spontaneously, genomic, epigenomic, and transcriptomic characteristics resembling advanced invasive LUSC are observed.²⁸ This process involves the simultaneous expression of immune checkpoint molecules and suppressive interleukins, raising hope for the possibility of intercepting progression to invasive cancer by inhibiting the blocking factors of anti-tumor immune response.

Adenocarcinoma carcinogenesis

Atypical adenomatous hyperplasia (AAH) is recognized as the only precursor of LUAD.²⁹ AAH may progress through preinvasive adenocarcinoma in situ (AIS), minimally invasive adenocarcinoma (MIA), and culminating in fully invasive LUAD (Fig.1).^{30,31,32,33} LUAD precursors present radiologically as ground-glass opacity (GGO) predominant pulmonary nodules. The detection of these nodules has increased due to the widespread adoption of low dose computed tomography (CT) in lung cancer screening and the use of high-resolution CT scans for various medical purposes.³⁴ Over the past decade, extensive analyses of AAH, AIS and MIA have revealed a progressive increase in the complexity of their molecular evolution and associated immune responses,^{35,36,37,38} with an increase in total mutation burden, copy number variant burden and cancer gene mutations in later-stage lesions.^{35,36,37,38} Canonical cancer gene mutations such as *KRAS*, *EGFR*, and *BRAF* appear to be early genomic events during LUAD carcinogenesis,^{35,36,37,38} in parallel with a gradual suppression of immunity, indicating ongoing “immunoediting”.^{37,38,39} Preclinical studies demonstrated that reprogramming the TME may promote anti-tumor immunity and reduce the burden of invasive adenocarcinoma in mouse models.⁴⁰ More recently, single-cell technologies have revealed the complexity of tumor evolution with unparalleled resolution and have pinpointed alveolar type 2 (AT2)-like cells as contributors to LUAD progression.^{41, 42} How the spatial context within the TME changes during different stages of disease progression remains to be clarified.

Biomarkers of early detection

Small nodules in lung parenchyma can be efficiently detected by low-dose CT to allow early diagnosis of lung cancer and reduction of lung cancer mortality.⁴³ Recent advances in this field included the application of radiomics and artificial intelligence (AI) to develop non-invasive biomarkers for lung cancer risk prediction, early detection and prognosis prediction.⁴⁴ Analysis of the airway transcriptome of minimally invasive collected bronchial and nasal epithelial cells has proven effective for early diagnosis of lung cancer.⁴⁵ Likewise, the detection and molecular profiling of circulating tumor cells (CTC) in blood has been the objective of many studies to identify CTC biomarkers, although their application in lung cancer screening was questioned due to reported low sensitivity in prospective studies.^{46,47} Cell-free nucleic acids (DNA and RNA), which can be passively and/or actively released by tumor cells and by other cells in the TME⁴⁸ have been the focus of many studies for early diagnosis, prognosis and prediction of therapy response. Low-pass whole genome sequencing of cell-free DNA (cfDNA) fragmentation profiles with the aid of AI has shown promising results in asymptomatic NSCLC.⁴⁹ Indeed, changes to the genomic and epigenetic architecture of cells during carcinogenesis result in a varied cfDNA

“fragmentome” in the peripheral blood.⁵⁰ Circulating cell-free microRNA in serum or plasma has also been investigated as a diagnostic tool and was validated in lung cancer screening trials.^{51, 52} More recently, a comprehensive proteomic analysis in plasma samples has provided biomarkers for early prediction of lung cancer.⁵³ Finally, the integration of multiple types of biomarkers (e.g. nucleic acids and proteins) has also been attempted to detect various cancer types, including lung cancer. While the specificity appeared high (>99%), the sensitivity was low (~20–30%), limiting its application as a first-line diagnostic tool in high-risk subjects.⁵⁴

Tumor heterogeneity

Heterogeneity is an inherent characteristic of all cancers, including NSCLC. It may refer to differences among tumors from different patients, known as intertumoral heterogeneity, which reflect distinct carcinogenic pathways in each tumor. Alternatively, it can describe variation within a single tumor, termed intratumoral heterogeneity (ITH). The latter involves the presence of multiple and diverse clonal cancer cell populations within a single tumor. This diversity underpins the evolution and remarkable adaptability of tumors to various environments, which may lead to progressive invasiveness and aggressiveness, contributing to treatment resistance. A greater understanding of ITH in NSCLC from molecular and histopathologic perspectives is cumulatively impacting patient clinical management (Fig.2).

Intratumor molecular heterogeneity

The coexistence of molecular and phenotypically distinct subclones within tumors are thought to develop under the so-called branched model of tumor evolution.⁵⁵ According to this model, tumors arise from a common ancestor eventually diverging and proliferating simultaneously, each with varying levels of efficacy. The punctuated model is a variant of the latter shaped by a macroevolution in which, after an extended stable period, many genomic aberrations occur within short bursts of time. Some tumors may evolve according to a neutral evolution model, in which cancer-driving alterations accumulate randomly within the cancer cell population without having a functional role in promoting tumor growth⁵⁶.

TRACKing Cancer Evolution through therapy [Rx] (TRACERx) is a prospective study which sought to define the role of genetic ITH on patient survival and disease characteristics.^{57,58} Inference of the clonal architecture in the tumors in the study demonstrated that cancer driver mutations are under positive selection, not only when they occur as truncal mutations, but also when present in tumor subclones.^{59,60} In the same study, a higher subclonal somatic copy number alteration (SCNA) burden, but not mutational burden, was associated with worse disease-free survival.⁵⁹ Additional biological consequences of genetic ITH include increasing neoantigen burden and subsequent immune response,⁶¹ in addition to the generation of drug resistance mutations.⁶²

Epigenetic alterations (e.g. DNA methylation and histone modifications) are crucial for tumor development and contribute to ITH. The most studied epigenetic deregulation in the context of ITH is DNA methylation. Extensive ITH in DNA promoter hypermethylation, driven by changes in DNA copy number, was found to contribute to the molecular and

phenotypic heterogeneity of LUAD and of lymph node metastases.⁶³ Further, genomic and DNA methylation changes appear to follow similar ITH evolutionary trajectories with a strong impact in cancer genes and pathways.⁶⁴ Given that drug development targeting epigenetic factors is underway, a deep understanding of epigenetic alterations in NSCLC will be essential for their effective implementation in the clinics.⁶⁵

ITH can also be observed at gene expression and protein levels, which does not always originate from genomic ITH.⁶⁶ Transcriptomic diversity has been found between primary tumors and metastasis and contributes to tumor progression.^{66,67} While multi-region sequencing provided foundational insights into ITH,^{68,35} advancements in single cell sequencing and spatial transcriptomics have elevated ITH research to a new level and have provided comprehensive datasets.⁶⁹

Intratumor spatial distribution of immune related proteins and patterns of proteins functionally involved in cell adhesion or endothelial cell interaction have been observed, some of which may be prognostic.^{70,71} Deciphering the link between the static cancer genotype and its dynamic transcription and protein expression offers novel multilayers insights on the mechanisms of cancer adaptation.^{72,73} The development of effective strategies to integrate ITH at the genomic, transcriptomic, and proteomic levels may provide opportunities for enhanced dynamic biological insights and the identification of novel therapeutic targets in lung cancer.^{74,75}

Histologic heterogeneity

The current World Health Organization (WHO) classification of lung cancers includes more than 20 entities, each defined by the unique histopathological appearances of the tumor cells and their growth pattern.⁷⁶ Morphological ITH is best observed in LUADs where the relative abundance of the different patterns of growth (lepidic, acinar, papillary, micropapillary and solid) is prognostic (Fig. 3).^{77,78} Lepidic patterns are considered *in situ* tumors and thus associate with good prognosis⁷⁹ while non-mucinous LUADs with predominantly micropapillary and solid growth patterns are associated with poor prognosis.⁸⁰ Based on the IASLC grading system, any tumor with predominant or $\geq 20\%$ of solid, micropapillary and complex glandular patterns are considered grade 3 (high grade).⁸¹ Importantly, previous gene-expression based subtypes of LUADs⁸² have not been correlated with the morphologic classification, possibly suggesting more complex multi-omics determinant of tumor growth patterns. The molecular mechanisms underlying morphological features of LUAD remain largely unknown, as do the molecular features associated with the development of lung cancers of mixed types.⁸³ A recent study employing micro-dissection of tumor regions in 19 LUADs with mixed histologic patterns or subtypes suggested an association of specific transcriptomic pathways with the different morphologies.⁸⁴ Other studies have reported that high-grade histologic patterns are associated with increased chromosomal complexity, higher burden of genomic aberrations and lower clonal diversity.^{85,86}

Clinical impact of tumor heterogeneity

Intertumoral heterogeneity and ITH also represent a major hurdle in the effectiveness of therapeutic strategies to treat lung cancer. Regarding intertumoral heterogeneity, even

within a defined histopathology, co-mutations and molecular differences among tumors can profoundly influence prognosis, tumor plasticity and therapeutic sensitivity.^{87,88,89} The concomitant *RB1* inactivation in *EGFR*-mutant LUADs may facilitate histopathological transformation in response to targeted therapy⁸⁸ and *TP53* mutations cooccurring with *EGFR* are more likely to induce mixed responses.⁹⁰ Similarly, *STK11* mutations are linked to reduced sensitivity to immune checkpoint inhibitors (ICIs) in *KRAS*-driven LUAD,⁹¹ while mutations in *KEAP1/NFE2L2* decrease sensitivity to different treatments and are associated with poorer prognosis.⁹² These data highlight the importance of determining molecular clonal heterogeneity in otherwise uniform NSCLC populations.

Radiomics has recently opened avenues for better imaging assessment of intertumoral heterogeneity. The visualization and assessment of the whole tumor as well as the metastatic sites for radiomic features using machine learning may become an effective prognostic tool for prediction of response to immunotherapies.⁹³ Radiomic data may also lead to opportunities to escalate or change treatment or consider multimodality treatment options.

Similarly, cfDNA has been used as another non-invasive alternative for tracking molecular heterogeneity and determining clinical management. Analysis of cfDNA in the TRACERx cohort over multiple timepoints showed that about 40% of patients had tumors with ITH, in which a dominant clone emerged and replaced others between surgical resection of their primary tumor and disease recurrence.⁹⁴

Tumor cell plasticity

Cellular lineage plasticity (LP) refers to the ability of cells to switch phenotypes to distinct developmental lineages; it is integral to processes such as embryogenesis, tissue repair, and homeostasis.⁹⁵ However, cancer cells may hijack these mechanisms to adapt to stimuli, or aid tumor progression and metastasis. LP is increasingly recognized as a resistance mechanism to targeted therapies in lung cancer. Notable lineage states in LUAD include cancer stem cell phenotypes, epithelial-mesenchymal transition states, histologic transformation to neuroendocrine characteristics often with small cell lung cancer (SCLC) histology, and transformation to LUSC phenotypes. The mechanisms of LP in lung cancer are still poorly understood, which pose therapeutic challenges in the management of NSCLC at high-risk of LP.

While there are distinctions between various forms of plasticity, such as EMT or adenoneuroendocrine and adeno-squamous transformation, there is substantial overlap in the features and mediators of these phenomena. These changes are generally associated with hybrid metastable states characterized by elevated cellular plasticity and stem-like features. Single cell RNA sequencing in a genetically engineered mouse models (GEMM) of LUAD demonstrated that longitudinal transcriptional heterogeneity with disease progression was stereotypic and reproducible.⁹⁶ Certain factors that govern cell fate decisions during development and organ formation have resurfaced in cancer biology as key regulators of ITH and LP. Lineage tracing studies have played a crucial role in mapping the connections between progenitor and differentiated cells, uncovering varying degrees of plasticity across different stages of differentiation. The presence of these cell states may predict for poorer survival and are more resistant to chemotherapy⁹⁷ and immunotherapy.⁹⁸

Histologic transformation

The histologic transformation of LUAD to SCLC or LUSC, with or without therapeutic pressure, epitomizes the LP observed in lung cancer.⁹⁹ This transformation process likely involves a complex interplay of genetic, transcriptomic, epigenetic and immune factors.^{100,101} The cell of origin, such as AT2 cells for LUAD, may play a key role in determining the predilection for certain oncogenic pathways. The bi-allelic inactivation of key tumor suppressor genes, *TP53* and *RB1*, is associated with histological transformation from LUAD to SCLC in tumors under tyrosine kinase inhibitors (TKIs) anti EGFR therapy.¹⁰² A ‘third hit’ such as FGF9 upregulation is associated with SCLC trans-differentiation.⁹⁹ However, the exact mechanisms remain unknown. Recently, through GEMM and single-cell RNA sequencing, the upregulation of transcriptional programs induced by expression of *Myc* in cooperation with *RB1* loss have been implicated.⁹⁶ In addition, an intermediate basal stem-like cell state induced by EGFR inhibition may facilitate a tolerance to Myc-driven histological transformation. Histological transformation of LUAD to LUSC, whereby de novo deficient *Stk11* triggers extracellular matrix remodelling and p63 upregulation,¹⁰³ may similarly be characterized by an intermediate cell state with predisposing genetic alterations governing lineage plasticity in response to therapy. In response to *KRAS* targeted therapy, concurrent *STK11* mutations may alter the regulation of chromatin accessibility. Consequently, modulation of lineage-related transcription factors and the ELF5-ΔNp63 axis may drive histological transformation.¹⁰⁴ Lastly, cell-extrinsic factors may also play a crucial role in shaping lineage cell states by providing a permissive microenvironment for histological transformation,^{105,106} although this remains underexplored to date.

Drug-tolerant persister cells

Residual cancer cells that persist during treatment serve as the reservoir for the emergence of drug-resistance. This has prompted research into the cellular and molecular basis of drug-tolerance, the evolution of drug tolerant persister (DTP) cells and the identification of therapeutic vulnerabilities to target them, mostly in the context of targeted therapies. In *EGFR*-driven LUAD patient-derived xenograft (PDX) models, the transcriptional profile of DTP cells to TKIs has been identified in subpopulations of treatment-naïve tumors suggesting that, in some cases, these cells are present prior to treatment.^{107,108} A role for epigenetic processes in maintaining the drug-tolerant state, including alterations in histone methylation patterns linked to repression of long interspersed repeat element 1 (LINE-1) elements was uncovered.^{109, 110} Additional mechanisms that lead to activation of NFκB and JAK-STAT pathways, the antioxidant KEAP1-NRF2 pathway, EMT, senescence, translation reprogramming, impaired cell death and alveolar regeneration contribute to reduced treatment sensitivity in DTP and constitute a common mechanism by which lung cancer cells withstand therapy-induced stress.^{111,112,113,114,115,92,116,107,117} Understanding when specific mechanisms of drug tolerance occur and how they can be targeted will be necessary to identify new approaches to mitigate or forestall the emergence of resistance.^{118,119,120,121}

Epithelial–mesenchymal transition (EMT)

EMT is a highly dynamic and reversible cellular program in which epithelial cells lose cell–cell junctions, display altered apical–basal polarity and acquire increased migratory capacity.¹²² EMT histologically may manifest in poorly differentiated NSCLC or sarcomatoid/spindle cell carcinoma. Recent efforts have contributed to uncovering the molecular mechanisms associated with EMT-mediated resistance. ZEB1 directly binds the promoter of BIM, a positive regulator of apoptosis, repressing its transcription.¹²³ Similarly, Aurora kinase (AURK) B mitigates BIM-induced apoptosis to promote cell survival to treatment.¹²⁴ Activation of the YAP/FOXM1 axis promotes increased abundance of spindle assembly checkpoint effectors and mediates EMT-associated EGFR inhibitor resistance.¹²⁵ Concordantly, model systems of osimertinib resistance caused by EMT display activation of ATR–CHK1–AURKB signaling.¹²⁴ EMT-mediated resistance has also been attributed to the reactivation of downstream signaling pathways, including sustained activation of AXL,^{126, 127,128} FGFR1^{129,130} and SRC.¹³¹ More recently CD70, which is highly expressed in NSCLC with mesenchymal phenotype,¹³² was reported to be a targetable mechanism of EMT-associated EGFR inhibitor resistance.¹³³ In depth molecular characterization of baseline, on-treatment and resistant tumors and circulating tumor cells, together with functional analyses, might also help capturing the diversity and dynamics of EMT and contribute to uncovering the molecular alterations underlying its role in resistance to therapy.¹²² These findings may reveal targetable dependencies and therapeutic combination rationales to re-sensitize mesenchymal lung cancers to targeted therapy in preclinical models.^{123,124,130,127,129,133}

Tumor microenvironment

Tumors, including NSCLC, consist of a diverse ecosystem of cells which, in addition to cancer cells, include a plethora of host cells with widely different and often opposing functions in oncogenesis. The importance of the immune cell contexture in NSCLC has been well established and draws from studies showing the TME to be strongly prognostic and the marked success of cancer immunotherapy increasingly becoming part of first-line care.^{134,75,135} Many of the principles governing the immune- and cancer cell interrelationship seem to be well preserved across NSCLC stages, histological and molecular subtypes with certain exceptions, further bolstering the broad use of immunotherapy clinically. However, durable benefits to cancer immunotherapy occur in a minority of NSCLC patients for reasons still incompletely understood, warranting a better understanding of tumor-immune cell dynamics in NSCLC to spark a new wave of effective therapeutic strategies resulting in long-lasting tumor control.

NSCLC immunogenicity and T-cell recognition

Intratumoral immune cells sculpt NSCLC genomic heterogeneity and vice versa, NSCLC cells influence immune cells through various mechanisms. Earlier findings in preclinical models showed increased tumorigenesis in the absence of key immune cells or anti-tumor effector molecules. This led to the understanding that tumors are constantly surveilled by cytotoxic T- and NK cells with immunogenic tumor cell clones being lysed in a process called ‘immuno-editing’. In the last decade, increasing evidence points to a role for T cells

in shaping tumor cell heterogeneity in NSCLC patients. Seminal studies including those from the TRACERx consortium have shown increased T-cell infiltration in patient tumors with high tumor-mutation burden (TMB), tumor areas exhibiting highly clonal neo-antigen expression, as opposed to low neo-antigen and clonally heterogeneous tumor areas.^{136,61} These cells, however, are found to be dysfunctional bearing an 'exhausted' phenotype characterized by the expression of multiple immune-regulatory molecules including LAG-3 and PD-1.^{61,137} Anti-PD-1 was especially effective in these patients and to a lesser extent in those with a more T-cell and neo-antigen devoid TME.^{136,61,137,138} Latter studies further showed compelling evidence of T-cells driving genomic tumor evolution, demonstrating decreased neo-antigen recognition potential in high T-cell infiltrated tumor regions through genomic, transcriptional and epigenetic mechanisms inducing immune-evasion (e.g. loss of HLA-alleles).^{139,140,141} Whereas in established tumors immune-evasion leads to further tumor outgrowth, earlier CIS lesion in LUSC that spontaneously regressed showed increased T-cell presence and inflammation.¹⁴² A significant proportion of CIS, however, still progresses to overt NSCLC exemplifying that immune evasion is a critical step early in tumorigenesis (Fig.4).^{26,142} These studies highlight the impact of T-cells and other immune cells in driving ITH following the recognition and killing of immunogenic tumor cells. This process in turn can be reinvigorated by ICI leading to potentially long-lasting and even complete responses in a subset of patients. Further research is needed to understand what determines tumor behavior, response to ICI, and to learn from cells in and outside the TME to further develop improved biomarkers and therapeutic strategies.

T-cell, myeloid and stromal cell heterogeneity in the TME and systemic anti-tumor immunity

NSCLC tumors can be roughly separated into T-cell inflamed tumors co-occurring with several different myeloid cells (e.g. dendritic cells, macrophages) and B-cells, and T-cell deserted or excluded tumors.^{143,75,135,144} The latter TME-contexture is more often seen in tumors from non/light-smokers, characterized by low TMB, loss of antigen-presentation machinery, strong driver mutations (e.g. EGFR and ALK) and can co-exist with a predominant neutrophilic infiltrate and/or stroma-rich regions with specific fibroblast-subsets mechanically impeding T-cell infiltration.^{145,146,147,148,135} However, not all intratumoral T-cells are indeed tumor-specific; varying but significant amounts of T-cells in the TME recognize viral antigens or other cancer-irrelevant peptides.^{149,150,151} Cancer-cell intrinsic mutations (e.g. in EGFR, STK11, genes involved in cell metabolism and increased TMB resulting in high neo-antigen load) and -extrinsic features (e.g. vasculature and associated hypoxia) are considered cardinal drivers of TME heterogeneity impacting the resultant anti-tumor immune response.¹⁵² Other oncogenes and tumor suppressor genes are known to affect the immune response. For example, MYC oncogenic activation can prevent an appropriate response to interferon-gamma (IFN γ).¹⁵³ Tumor-specific genetic alterations that impact the processing and function of the HLA-I complex (e.g., B2M, TAP1) or the response to IFN γ (e.g., JAK2, IFNGR1) are also common in lung cancer, influencing the composition of the TME^{147,154} and impairing the response to ICIs.¹⁴⁶

Crucial for induction of anti-tumor T-cells and efficacy of ICI is effective priming in tumor-draining lymph nodes (TDLNs) by dendritic cells presenting tumor antigens^{155,156}. Priming

is an ongoing, dynamic process allowing for the continuous seeding and replenishment of T-cells also in response to ICI.^{157,150} The importance of this process extends beyond immunity to the primary tumor and involves a systemic immune surveillance program potentially limiting the occurrence of distant metastasis.^{158,159} Importantly, the initiation and strength of this systemic anti-tumor immune response can be severely hampered by local immune suppression in TDLNs by e.g. regulatory Tregs^{160,161} or dendritic cells.¹⁶² Moreover, TDLN-like structures in the TME called mature tertiary lymphoid structures (TLS) are also identified in NSCLC and correlate with an immune inflammatory environment and favorable prognosis, potentially by regulating T-cell attraction and maturation in concert with other perivascular niches.^{163,164} Future strategies aimed at sensitizing NSCLC patients, and prolonging responses to ICI, will have to consider aspects regarding systemic anti-tumor immunity.

Translational aspects to ICI resistance

Contrary to earlier beliefs, the response to ICI in NSCLC seems to be not only dependent on activation of pre-existing tumor T-cells but relies on a replenishment by circulating T-cells including T-cells with early differentiation phenotypes and novel TCR-specificities.^{157, 158, 159} Primary resistance to ICI can occur at multiple steps in anti-tumor immunity initiation ranging from a paucity of immunogenic tumor antigens, antigen presentation, strong immune suppression or high stromal/vascular impediment, as highlighted earlier. A significant proportion of patients, however, respond initially but eventually progress in a process called secondary immune resistance. Tumor behavior in secondary resistance shares similarities to primary immune resistance, but there are important differences as well including the frequent occurrence of oligoprogression (progression of a single lesion, with persistent immune control of others) and related to this, better survival as compared to primary resistance.¹⁶⁵ Genomic and proteomic characterization of the TME of progressing lesions has identified similar phenomena as observed in primary immune resistance relating to impaired immunorecognition or antigen presentation contribute to immune evasion (e.g. decreased expression of the HLA-protein complex,^{166, 146} loss of clonal neo-antigens and acquisition/selection of mutations associated with ICI-resistance (e.g. in *STK11*).^{91,167,165} Additionally, chronic IFN signaling in response to ICI may paradoxically induce PD-L1-independent routes of immune suppression through epigenetic alterations induce inhibitor ligand and molecule expression capable of suppressing more exhaustion-sensitive CD8⁺ T-cells.^{165,168} Interestingly, blocking (chronic) IFN-signaling with JAK-inhibitors has shown potential together with ICI in NSCLC.¹⁶⁹ These studies and others highlight the importance of comprehensive multi-omic and translational studies of NSCLC resistant to ICI to develop rational immunotherapy combinations.

Metabolic Landscape of Lung Cancer

The metabolic reprogramming of NSCLC has been studied in recent years with the aim of identifying vulnerabilities that could be translated into novel therapeutic approaches (Fig.5). Several studies have highlighted the role of fatty acid, glucose, and mitochondrial metabolisms in NSCLC tumorigenesis. The RAS pathway is important in metabolic rewiring and potentially targetable in NSCLC. KRAS-mutant NSCLCs with co-mutations in STK11

and KEAP1 represent a distinct subset of the disease with unique characteristics and treatment challenges.^{170, 171}

In NSCLC, glutathione, an essential antioxidant in cells, and associated enzymes are often not adequately regulated.^{172,173} This promotes resistance to oxidative stress and helps cancer cells survive and multiply.¹⁷³ STK11 is widely studied as the kinase that activates AMPK to rewire metabolism by modulating mTOR signaling.^{174,175} Recent studies have shown that the salt-inducible kinase (SIK) family members (SIK1 and SIK3) mediate the tumor suppressor role of STK11 in NSCLC but not AMPK.^{176,177} KEAP1 is a key regulator of the cellular antioxidant response. It controls the activity of NRF2, a transcription factor that regulates gene expression in antioxidant and detoxification processes. The KEAP1-NRF2 pathway regulates the expression of antioxidant genes, which promotes cancer cell survival and resistance to therapy.¹⁷⁸ The interplay between glutathione and the KEAP1-NRF2 pathway in NSCLC underscores the importance of understanding redox regulation in cancer progression and treatment. KRAS-mutant NSCLC with co-mutations in STK11 and KEAP1 upregulates ferroptosis evasion pathways. Targeting ferroptosis evasion by inhibiting stearoyl CoA desaturase 1 (SCD1), a master regulator of fatty acid metabolism, has shown promise in preclinical models.¹⁷¹ SCD1 can directly regulate the SIK1, which disrupts the multimodal ferroptosis protection of STK11/KEAP1 co-mutated NSCLC cells.^{171,179,177} Studies have shown that the SIK1-NRF2 axis can modulate the sensitivity of STK11/KEAP1 co-mutant cancer cells to ferroptotic cell death and chemotherapy.^{176,177} SIK1 also controls the serine glycine one-carbon metabolism (SGOC), which plays a crucial role in nucleotide synthesis, methylation reactions, and redox balance within the cell. Most interestingly, SIK1 was found to negatively regulate the SGOC in STK11/KEAP1 NSCLCs by activating SHMT, which increases the cellular antioxidant mechanism.¹⁸⁰ Therefore, further exploration of the interplay between SIK1, NRF2, and the STK11/KEAP1 co-mutant genotype holds promise for identifying potential therapeutic targets and developing personalized treatment strategies for patients with oncogene-driven subsets of NSCLC.

Metabolic reprogramming may facilitate the formation of an immunosuppressive microenvironment by tumor cells, where lactic acid, a metabolite of glycolysis, is now documented to contribute to this process.^{181,182} Lactic acid can directly suppress the TME by inducing apoptosis of natural killer cells (NK) and natural killer T-cells (NKT).^{183,184} In addition, it can inhibit the proliferation of cytotoxic lymphocytes,¹⁸⁵ block IFN γ and IL4 production by NKT cells, inhibit the differentiation of dendritic cells (DCs) and increase the production of IL10. Furthermore, lactic acid has been shown to confer changes in the innate and adaptive immune responses by driving the development of myeloid-derived suppressor cells.¹⁸⁶ Studies have demonstrated a positive correlation of high lactic acid concentrations in the TME with tumor invasion and metastasis in multiple cancer types including lung cancer.^{187,188,189} As such, targeting tumor cell production and transport of lactic acid may provide an effective strategy for anti-tumor therapy.¹⁹⁰

Considering the immunosuppressive effects of lactate metabolism in lung cancer and its effects on anti-tumor immunity, several signaling pathways and targets have been reported that may offer therapeutic targeting or inhibition. Monocarboxylate transporters^{181,182,183,184} are lactic acid transporters¹⁹¹ and the efflux of lactate from tumor cells

is mediated by monocarboxylate transporter 4 (MCT4), also known as SLC16A3, while SLC16A1 is thought to mediate its influx.¹⁹² While the role of SLC16A3 in tumor development and lactate metabolism is not well understood, studies have reported that SLC16A3 is highly expressed in lung cancer¹⁹³ and is negatively correlated with immune cell infiltration. Pharmacological inhibition of SLC16A3 in tumor cells attenuated intercellular lactic acid exchange, resulting in the reduction of acidity within the TME and increased the efficacy of anti-PD-1 therapy, highlighting the potential targeting of lactate acid transporters in overcoming immunotherapy resistance.¹⁸¹ A number of MCT1 inhibitors have been reported that have shown promising pre-clinical activity and include SR138009¹⁹⁴ and AZD3965.¹⁹⁵ The process of lactylation has recently emerged as an important form of protein modification where studies have demonstrated that lactate modulates cell metabolism in NSCLC through increased histone lactylation in the promoter regions of the HK-1 and IDH3G genes.¹⁹⁶

Lactate has also been implicated as a messenger in the cross talk between cancer cells and immune cells, in particular, tumor-associated macrophages, where it has been shown to influence macrophage polarization.¹⁹⁷ Furthermore, the nuclear factor of activated T-cell c2 (NFATc2) promotes M2 macrophage polarization by regulating USP17 via transcriptional stimulation. Upon overexpression of USP17 in LUAD cell lines, lactate levels were upregulated, an effect that was reversed in response to inhibition of NFAT.¹⁹⁸

More recently, there has been an emerging interest in the metabolic reprogramming of NSCLC by oncogenic driver mutations such as EGFR, EML4-ALK and BRAF.¹⁹⁹ Research examining the metabolic alterations in EGFR mutant NSCLC highlights the dependency of these tumors on glucose,²⁰⁰ glutamine²⁰¹ and lactate metabolism,²⁰² in addition to fatty acid synthesis.²⁰³ Compared to EGFR wild-type and EGFR-mutated NSCLC, patients with ALK rearrangements exhibit higher glucose metabolism, where targeting of EML4-ALK fusions with ceritinib in xenograft models demonstrates a reduction in glycolysis. It has been suggested that this process is mediated through the EML4-ALK-HIF1 α -HK2 axis.²⁰⁴ While few studies have been reported on the metabolic effects of BRAF mutations in NSCLC, it has been shown that BRAF V600E-driven tumors can maintain mitochondrial function and glutamine metabolism through their addiction to autophagy.²⁰⁵ Furthermore, the failure of late stage Atg7-deficient tumors to thrive was a consequence of impaired mitochondrial function and limited availability of substrates such as glutamine, resulting in tumor crisis which was incompatible with tumor growth.²⁰⁶

Further studies to better understand the metabolic plasticity that exists between tumor cells and immune cells may provide new knowledge supporting the development of novel combination (metabolic and immune) therapies in lung cancer.

Microbiome

Although the lungs were previously considered sterile organs, despite constantly being exposed to microorganisms present in the upper respiratory airways and in airborne bioaerosols, with the advent of NGS, a variety of lung commensal microbiota were identified.^{207, 208} Lung microbiota interacts with the gut microbiota (Gut-Lung axis) by

means of ligands and metabolites, which might be pivotal to train the lung immune system and protect from exacerbated inflammatory responses to inhaled antigens.²⁰⁹ This commensal microbiota tolerance develops early in the newborns (within ~2 months from birth)²¹⁰ and appears to play important roles in preventing inflammation-related lung damage as well as lung carcinogenesis.

Microbiome in NSCLC initiation and progression

Dysregulation of lung microbiota might lead to changes in the pre-neoplastic/neoplastic lung microenvironment by reshaping tumor-infiltrating immune cells, ultimately contributing to cancer onset and progression.^{211,212} This may be due to the role of chronic inflammation in lung carcinogenesis, with evidence that repeated exposure to antibiotics was associated to NSCLC risk^{213,214}. Several bacteria have been associated with chronic inflammation and subsequent increased cancer risk including lung cancer, like *Mycobacterium tuberculosis*.²¹⁵ Liu *et al.*²¹⁶ showed that lung microbiota in patients with NSCLC are enriched in genus *Streptococcus* while depleted in genus *Staphylococcus*. It was also reported that genus *Thermus* was more abundant in patients with advanced NSCLC, while genus *Legionella* was more abundant in patients who develop metastases.²¹⁷ Ma *et al.* found that NSCLC patients had enrichment of Th1 and Th17 cells reacting to *Streptococcus salivarius* and *Streptococcus agalactiae* compared to healthy controls.²¹⁸

Microbiome in lung cancer therapies

The therapeutic efficacy of ICIs has been correlated with the composition of the microbiome, in combination with the effect on the host immune system. Routy *et al.*²¹⁹ observed an increased relative abundance of *Akkermansia muciniphila* in anti-PD-1 responders compared to non-responders. However, taxonomic-centric microbiome studies have shown huge limitations, as different immune-regulatory strains with distinct taxonomic assignments within the same cancer types receiving the same treatment have been reported.²²⁰ This discrepancy is likely due to the rapid evolution of bacteria at the strain level and the profound inter-individual heterogeneity of the human microbiota, in addition to differences in microbiota acquisition, processing and identification. Assessing the functional capacity of the microbiota across different patient cohorts using standardized approaches may serve as a better biomarker in this setting.

Future perspectives

Basic and translational research in lung cancer during the last three decades has greatly advanced our understanding of the molecular pathogenesis and biology of lung cancer. Technological advances in high throughput nucleic acid sequencing and proteomic analysis, computational science, medicinal chemistry, molecular modeling, biobanking and large data science have accelerated these research achievements, with transformational impacts in the clinical detection, diagnosis, treatment and management of patients with lung cancer. Continued advances in technologies, including single cell multi-omics analyses and AI will continue to accelerate our research advances, raising further optimism on the future prospect of curing lung cancer in a larger number of patients. In addition, for these innovations to translate into meaningful clinical impact, integrating patient-derived samples, including pre-

and post-treatment, multi-region samples in well-annotated patient cohorts will not only refine our understanding of NSCLC biology but also accelerate the development of truly personalized treatment strategies, ensuring these advancements reach the patients who need them most.

In the field of early-stage lung cancer, efforts have focused on monitoring high-risk groups, but there is a need for solutions in biomarker screening for early detection, preventive treatments for cancer interception, and risk stratification to avoid overtreatment. Developing standardized immune biomarkers for pre-cancerous conditions is crucial. Additionally, integrating AI-assisted medical decision-making would provide guidance on clinical decisions within the disease's temporal context. Leveraging the latest research findings requires conducting prospective, multicentric clinical trials on biomarkers for detection, prediction, and interception. Although it has long been recognized that LUAD precursors often manifest as GGO-predominant lung nodules, and that severe dysplasia and CIS serve as LUSC precursors, efforts to intercept these lung cancer precursors have been limited by our rudimentary understanding of the molecular events and tumor microenvironment changes driving malignant transformation and neoplastic evolution. However, emerging evidence highlighting the simpler molecular landscape and more active immunity in lung cancer precursors has spurred interest in immune interception, aiming to leverage the enhanced immune activity in precancers to prevent the development of invasive lung cancers. In this context, multiple clinical trials that explore the efficacy of immunotherapies ([NCT04789681](#), [NCT03634241](#), [NCT03347838](#)) have been initiated and encouraging results would represent a significant step forward in immune interception for lung cancer prevention.

Tumor heterogeneity is a fundamental characteristic of neoplastic progression including invasion, tumor growth and metastasis. It also plays a role in response to initial therapy and the development of resistance mechanisms to therapy. The TRACERx studies have provided some clues to the nature and mechanism of tumor heterogeneity in lung cancer, yet these are likely to represent the tip of an iceberg, as to date, analyses have mainly been limited to exome, gene copy number and transcriptome levels. Further functional genomics analyses are also strongly recommended. Heterogeneity at the histological level is common involving all types of NSCLC, yet the molecular basis of such heterogeneity is poorly understood. Future studies will need to be ambitious and involve multiple 'omics analyses, single cell and spatial multi-omics analyses, as well as samples from primary versus metastatic tumors, and pre- versus post-treatment tumors. The incorporation of living patient-derived cell lines, organoids and xenografts also provides great benefits in the ability to model tumor biology with functional studies.

The development of resistance is almost a universal feature of systemic therapies in lung cancer, and despite extensive research, limited progress has been achieved in understanding the mechanisms and strategies to overcome them. In the field of targeted therapies, recent research interest has focused on the mechanisms of drug tolerance and plasticity, the latter involving histologic transformation from adenocarcinoma to neuroendocrine or squamous cell carcinoma. Future research to understand the mechanism of DTPs may lead to novel strategies to improve treatment efficacy and cure rates for these targeted

therapies. Understanding the mechanisms of plasticity may lead to new strategies to identify predictive biomarkers for their development and/or mitigate resistance-causing histologic transformation.

Advancing our understanding of lineage plasticity in lung cancer is critical for overcoming therapeutic resistance and improving outcomes. Future research must prioritize deciphering the molecular drivers of LP, such as epigenetic reprogramming, TME influences, and key transcriptional regulators like ZEB1 and YAP/FOXM1. Targeting EMT-associated dependencies, immune evasion pathways, and plastic cancer stem cells offers promising avenues for intervention. Integrating single-cell profiling and functional studies with clinical insights will drive personalized therapies addressing LP-driven resistance mechanisms.

A large subset of patients does not respond to currently approved or experimental ICIs, most likely due to limited tumor immunogenicity and T-cell presence in the TME. To improve response rates, future studies will have to shift from targeting pre-existing exhausted tumor-infiltrating T cells to strategies incorporating insights from early T-cell priming, homing and systemic immunosurveillance, to co-opting cells in the TME (e.g. macrophages, fibroblasts) or genetically modifying T cells as cellular therapies. Linking tumor cell biology (including oncogene-driven metabolic vulnerabilities, such as ferroptosis evasion, redox imbalance, and lactate generation) to immune contexture and downstream T-cell responses will be critical to reverse the T-cell hostile environment in NSCLC. Finally, whereas tumor vaccination seems of limited utility in the advanced stage disease setting, pivoting to earlier (including preinvasive) and neo-adjuvant stages together with ICI will be critical for durable memory and resulting clinical responses.

Current technologies enabling multi-omics analyses, combined with AI, have recently led to exciting discoveries in translational research. These advancements will further accelerate our knowledge and understanding of NSCLC biology, offering hope for less toxic and more personalized therapeutic strategies both in the frontline and at relapse. These innovations bring the promise of extended survival for many patients and the potential to increase the chance of cure for a greater number of patients.

This review is based on the knowledge of members of the IASLC Basic and Translational Research Committee, thus its scope may be limited by their expertise. In the future, additional more focused and in-depth reviews will be published by the committee.

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References

1. Bosse Y, Amos CI. A Decade of GWAS Results in Lung Cancer. *Cancer Epidemiol Biomarkers Prev* 2018;27:363–379. [PubMed: 28615365]

2. Panagiotou E, Vathiotis IA, Makrythanasis P, et al. Biological and therapeutic implications of the cancer-related germline mutation landscape in lung cancer. *Lancet Respir Med* 2024;12:997–1005. [PubMed: 38885686]
3. Mukherjee S, Bandlamudi C, Hellmann MD, et al. Germline Pathogenic Variants Impact Clinicopathology of Advanced Lung Cancer. *Cancer Epidemiol Biomarkers Prev* 2022;31:1450–1459. [PubMed: 35477182]
4. Liu M, Liu X, Suo P, et al. The contribution of hereditary cancer-related germline mutations to lung cancer susceptibility. *Transl Lung Cancer Res* 2020;9:646–658. [PubMed: 32676327]
5. Sorscher S, LoPiccolo J, Heald B, et al. Rate of Pathogenic Germline Variants in Patients With Lung Cancer. *JCO Precis Oncol* 2023;7:e2300190. [PubMed: 37992258]
6. Byun J, Han Y, Li Y, et al. Cross-ancestry genome-wide meta-analysis of 61,047 cases and 947,237 controls identifies new susceptibility loci contributing to lung cancer. *Nat Genet* 2022;54:1167–1177. [PubMed: 35915169]
7. Gorman BR, Ji SG, Francis M, et al. Multi-ancestry GWAS meta-analyses of lung cancer reveal susceptibility loci and elucidate smoking-independent genetic risk. *Nat Commun* 2024;15:8629. [PubMed: 39366959]
8. Dai J, Lv J, Zhu M, et al. Identification of risk loci and a polygenic risk score for lung cancer: a large-scale prospective cohort study in Chinese populations. *Lancet Respir Med* 2019;7:881–891. [PubMed: 31326317]
9. Lebrecht MB, Smith MJ, Crosbie EJ, et al. Validation of lung cancer polygenic risk scores in a high-risk case-control cohort. *Genet Med* 2023;25:100882. [PubMed: 37154150]
10. Boumtje V, Manikpurage HD, Li Z, et al. Polygenic inheritance and its interplay with smoking history in predicting lung cancer diagnosis: a French-Canadian case-control cohort. *EBioMedicine* 2024;106:105234. [PubMed: 38970920]
11. Zhang P, Chen PL, Li ZH, et al. Association of smoking and polygenic risk with the incidence of lung cancer: a prospective cohort study. *Br J Cancer* 2022;126:1637–1646. [PubMed: 35194190]
12. Zhu M, Lv J, Huang Y, et al. Ethnic differences of genetic risk and smoking in lung cancer: two prospective cohort studies. *Int J Epidemiol* 2023;52:1815–1825. [PubMed: 37676847]
13. Shi J, Shiraishi K, Choi J, et al. Genome-wide association study of lung adenocarcinoma in East Asia and comparison with a European population. *Nat Commun* 2023;14:3043. [PubMed: 37236969]
14. Duncan MS, Diaz-Zabala H, Jaworski J, et al. Interaction between Continuous Pack-Years Smoked and Polygenic Risk Score on Lung Cancer Risk: Prospective Results from the Framingham Heart Study. *Cancer Epidemiol Biomarkers Prev* 2024;33:500–508. [PubMed: 38227004]
15. Hung RJ, Warkentin MT, Brhane Y, et al. Assessing Lung Cancer Absolute Risk Trajectory Based on a Polygenic Risk Model. *Cancer Res* 2021;81:1607–1615. [PubMed: 33472890]
16. Osarogiabon RU, Yang PC, Sequist LV. Expanding the Reach and Grasp of Lung Cancer Screening. *Am Soc Clin Oncol Educ Book* 2023;43:e389958. [PubMed: 37098234]
17. Chang GC, Chiu CH, Yu CJ, et al. Low-dose CT screening among never-smokers with or without a family history of lung cancer in Taiwan: a prospective cohort study. *Lancet Respir Med* 2024;12:141–152. [PubMed: 38042167]
18. Masquelier B, Breilh D, Neau D, et al. Human immunodeficiency virus type 1 genotypic and pharmacokinetic determinants of the virological response to lopinavir-ritonavir-containing therapy in protease inhibitor-experienced patients. *Antimicrob Agents Chemother* 2002;46:2926–2932. [PubMed: 12183249]
19. O’Keeffe LM, Taylor G, Huxley RR, et al. Smoking as a risk factor for lung cancer in women and men: a systematic review and meta-analysis. *BMJ Open* 2018;8:e021611.
20. Landrigan PJ, Fuller R, Acosta NJR, et al. The Lancet Commission on pollution and health. *Lancet* 2018;391:462–512. [PubMed: 29056410]
21. Qi C, Sun SW, Xiong XZ. From COPD to Lung Cancer: Mechanisms Linking, Diagnosis, Treatment, and Prognosis. *Int J Chron Obstruct Pulmon Dis* 2022;17:2603–2621. [PubMed: 36274992]
22. Hill W, Lim EL, Weeden CE, et al. Lung adenocarcinoma promotion by air pollutants. *Nature* 2023;616:159–167. [PubMed: 37020004]

23. Wang JH, Pfeiffer RM, Musgrove D, et al. Cancer Mortality Among Solid Organ Transplant Recipients in the United States During 1987–2018. *Transplantation* 2023;107:2433–2442. [PubMed: 37291711]
24. Krishna C, Tervi A, Saffern M, et al. An immunogenetic basis for lung cancer risk. *Science* 2024;383:ead3808. [PubMed: 38386728]
25. Mong C, Garon EB, Fuller C, et al. High prevalence of lung cancer in a surgical cohort of lung cancer patients a decade after smoking cessation. *J Cardiothorac Surg* 2011;6:19. [PubMed: 21352550]
26. Mascaux C, Angelova M, Vasaturo A, et al. Immune evasion before tumour invasion in early lung squamous carcinogenesis. *Nature* 2019;571:570–575. [PubMed: 31243362]
27. Beane JE, Mazzilli SA, Campbell JD, et al. Molecular subtyping reveals immune alterations associated with progression of bronchial premalignant lesions. *Nat Commun* 2019;10:1856. [PubMed: 31015447]
28. Teixeira VH, Pipinikas CP, Pennycuik A, et al. Deciphering the genomic, epigenomic, and transcriptomic landscapes of pre-invasive lung cancer lesions. *Nat Med* 2019;25:517–525. [PubMed: 30664780]
29. McGuire S World Cancer Report 2014. Geneva, Switzerland: World Health Organization, International Agency for Research on Cancer, WHO Press, 2015. *Adv Nutr* 2016;7:418–419. [PubMed: 26980827]
30. Detterbeck FC, Homer RJ. Approach to the ground-glass nodule. *Clin Chest Med* 2011;32:799–810. [PubMed: 22054887]
31. Kodama K, Higashiyama M, Takami K, et al. Treatment strategy for patients with small peripheral lung lesion(s): intermediate-term results of prospective study. *Eur J Cardiothorac Surg* 2008;34:1068–1074. [PubMed: 18760618]
32. Mun M, Kohno T. Efficacy of thoracoscopic resection for multifocal bronchioloalveolar carcinoma showing pure ground-glass opacities of 20 mm or less in diameter. *J Thorac Cardiovasc Surg* 2007;134:877–882. [PubMed: 17903500]
33. Ohtsuka T, Watanabe K, Kaji M, et al. A clinicopathological study of resected pulmonary nodules with focal pure ground-glass opacity. *Eur J Cardiothorac Surg* 2006;30:160–163. [PubMed: 16723239]
34. National Lung Screening Trial Research T, Aberle DR, Adams AM, et al. Reduced lung-cancer mortality with low-dose computed tomographic screening. *N Engl J Med* 2011;365:395–409. [PubMed: 21714641]
35. Hu X, Fujimoto J, Ying L, et al. Multi-region exome sequencing reveals genomic evolution from preneoplasia to lung adenocarcinoma. *Nat Commun* 2019;10:2978. [PubMed: 31278276]
36. Hu X, Estecio MR, Chen R, et al. Evolution of DNA methylome from precancerous lesions to invasive lung adenocarcinomas. *Nat Commun* 2021;12:687. [PubMed: 33514726]
37. Zhang C, Zhang J, Xu FP, et al. Genomic Landscape and Immune Microenvironment Features of Preinvasive and Early Invasive Lung Adenocarcinoma. *J Thorac Oncol* 2019;14:1912–1923. [PubMed: 31446140]
38. Chen K, Bai J, Reuben A, et al. Multiomics Analysis Reveals Distinct Immunogenomic Features of Lung Cancer with Ground-Glass Opacity. *Am J Respir Crit Care Med* 2021;204:1180–1192. [PubMed: 34473939]
39. Dejima H, Hu X, Chen R, et al. Immune evolution from preneoplasia to invasive lung adenocarcinomas and underlying molecular features. *Nat Commun* 2021;12:2722. [PubMed: 33976164]
40. Yuan B, Clowers MJ, Velasco WV, et al. Targeting IL-1beta as an immunopreventive and therapeutic modality for K-ras-mutant lung cancer. *JCI Insight* 2022;7.
41. Wang Z, Li Z, Zhou K, et al. Deciphering cell lineage specification of human lung adenocarcinoma with single-cell RNA sequencing. *Nat Commun* 2021;12:6500. [PubMed: 34764257]
42. Yang D, Jones MG, Naranjo S, et al. Lineage tracing reveals the phylodynamics, plasticity, and paths of tumor evolution. *Cell* 2022;185:1905–1923 e1925. [PubMed: 35523183]
43. de Koning HJ, van der Aalst CM, de Jong PA, et al. Reduced Lung-Cancer Mortality with Volume CT Screening in a Randomized Trial. *N Engl J Med* 2020;382:503–513. [PubMed: 31995683]

44. Tunalı I, Gillies RJ, Schabath MB. Application of Radiomics and Artificial Intelligence for Lung Cancer Precision Medicine. *Cold Spring Harb Perspect Med* 2021;11.
45. Team AS. Shared Gene Expression Alterations in Nasal and Bronchial Epithelium for Lung Cancer Detection. *J Natl Cancer Inst* 2017;109.
46. Gallo M, De Luca A, Maiello MR, et al. Clinical utility of circulating tumor cells in patients with non-small-cell lung cancer. *Transl Lung Cancer Res* 2017;6:486–498. [PubMed: 28904891]
47. Godschalk MF, Downs RW. Effect of short-term glucocorticoids on serum osteocalcin in healthy young men. *J Bone Miner Res* 1988;3:113–115. [PubMed: 3264991]
48. Stejskal P, Goodarzi H, Srovnal J, et al. Circulating tumor nucleic acids: biology, release mechanisms, and clinical relevance. *Mol Cancer* 2023;22:15. [PubMed: 36681803]
49. Mazzone PJ, Bach PB, Carey J, et al. Clinical Validation of a Cell-Free DNA Fragmentome Assay for Augmentation of Lung Cancer Early Detection. *Cancer Discov* 2024;14:2224–2242. [PubMed: 38829053]
50. Cristiano S, Leal A, Phallen J, et al. Genome-wide cell-free DNA fragmentation in patients with cancer. *Nature* 2019;570:385–389. [PubMed: 31142840]
51. Montani F, Marzi MJ, Dezi F, et al. miR-Test: a blood test for lung cancer early detection. *J Natl Cancer Inst* 2015;107:djv063. [PubMed: 25794889]
52. Pastorino U, Boeri M, Sestini S, et al. Baseline computed tomography screening and blood microRNA predict lung cancer risk and define adequate intervals in the BioMILD trial. *Ann Oncol* 2022;33:395–405. [PubMed: 35091076]
53. Davies MPA, Sato T, Ashoor H, et al. Plasma protein biomarkers for early prediction of lung cancer. *EBioMedicine* 2023;93:104686. [PubMed: 37379654]
54. Lennon AM, Buchanan AH, Kinde I, et al. Feasibility of blood testing combined with PET-CT to screen for cancer and guide intervention. *Science* 2020;369.
55. Gerlinger M, Rowan AJ, Horswell S, et al. Intratumor heterogeneity and branched evolution revealed by multiregion sequencing. *N Engl J Med* 2012;366:883–892. [PubMed: 22397650]
56. Ashouri A, Zhang C, Gaiti F. Decoding Cancer Evolution: Integrating Genetic and Non-Genetic Insights. *Genes (Basel)* 2023;14.
57. Jamal-Hanjani M, Hackshaw A, Ngai Y, et al. Tracking genomic cancer evolution for precision medicine: the lung TRACERx study. *PLoS Biol* 2014;12:e1001906. [PubMed: 25003521]
58. Frankell AM, Dietzen M, Al Bakir M, et al. Author Correction: The evolution of lung cancer and impact of subclonal selection in TRACERx. *Nature* 2024;631:E15. [PubMed: 38965439]
59. Frankell AM, Dietzen M, Al Bakir M, et al. The evolution of lung cancer and impact of subclonal selection in TRACERx. *Nature* 2023;616:525–533. [PubMed: 37046096]
60. Medina PP, Romero OA, Kohno T, et al. Frequent BRG1/SMARCA4-inactivating mutations in human lung cancer cell lines. *Hum Mutat* 2008;29:617–622. [PubMed: 18386774]
61. McGranahan N, Furness AJ, Rosenthal R, et al. Clonal neoantigens elicit T cell immunoreactivity and sensitivity to immune checkpoint blockade. *Science* 2016;351:1463–1469. [PubMed: 26940869]
62. Isozaki H, Sakhtemani R, Abbasi A, et al. Therapy-induced APOBEC3A drives evolution of persistent cancer cells. *Nature* 2023;620:393–401. [PubMed: 37407818]
63. Dietz S, Lifshitz A, Kazdal D, et al. Global DNA methylation reflects spatial heterogeneity and molecular evolution of lung adenocarcinomas. *Int J Cancer* 2019;144:1061–1072. [PubMed: 30350867]
64. Hua X, Zhao W, Pesatori AC, et al. Genetic and epigenetic intratumor heterogeneity impacts prognosis of lung adenocarcinoma. *Nat Commun* 2020;11:2459. [PubMed: 32424208]
65. Chao YL, Pecot CV. Targeting Epigenetics in Lung Cancer. *Cold Spring Harb Perspect Med* 2021;11.
66. Martinez-Ruiz C, Black JRM, Puttick C, et al. Genomic-transcriptomic evolution in lung cancer and metastasis. *Nature* 2023;616:543–552. [PubMed: 37046093]
67. Lee WC, Diao L, Wang J, et al. Multiregion gene expression profiling reveals heterogeneity in molecular subtypes and immunotherapy response signatures in lung cancer. *Mod Pathol* 2018;31:947–955. [PubMed: 29410488]

68. Zhang J, Fujimoto J, Zhang J, et al. Intratumor heterogeneity in localized lung adenocarcinomas delineated by multiregion sequencing. *Science* 2014;346:256–259. [PubMed: 25301631]
69. Gavish A, Tyler M, Greenwald AC, et al. Hallmarks of transcriptional intratumour heterogeneity across a thousand tumours. *Nature* 2023;618:598–606. [PubMed: 37258682]
70. Soltis AR, Bateman NW, Liu J, et al. Proteogenomic analysis of lung adenocarcinoma reveals tumor heterogeneity, survival determinants, and therapeutically relevant pathways. *Cell Rep Med* 2022;3:100819. [PubMed: 36384096]
71. Wu HJ, Temko D, Maliga Z, et al. Spatial intra-tumor heterogeneity is associated with survival of lung adenocarcinoma patients. *Cell Genom* 2022;2.
72. Gillette MA, Satpathy S, Cao S, et al. Proteogenomic Characterization Reveals Therapeutic Vulnerabilities in Lung Adenocarcinoma. *Cell* 2020;182:200–225 e235. [PubMed: 32649874]
73. Sharpnack MF, Ranbaduge N, Srivastava A, et al. Proteogenomic Analysis of Surgically Resected Lung Adenocarcinoma. *J Thorac Oncol* 2018;13:1519–1529. [PubMed: 30017829]
74. Mund A, Coscia F, Kriston A, et al. Deep Visual Proteomics defines single-cell identity and heterogeneity. *Nat Biotechnol* 2022;40:1231–1240. [PubMed: 35590073]
75. Sorin M, Rezanejad M, Karimi E, et al. Single-cell spatial landscapes of the lung tumour immune microenvironment. *Nature* 2023;614:548–554. [PubMed: 36725934]
76. Nicholson AG, Tsao MS, Beasley MB, et al. The 2021 WHO Classification of Lung Tumors: Impact of Advances Since 2015. *J Thorac Oncol* 2022;17:362–387. [PubMed: 34808341]
77. Travis WD, Brambilla E, Noguchi M, et al. International association for the study of lung cancer/american thoracic society/european respiratory society international multidisciplinary classification of lung adenocarcinoma. *J Thorac Oncol* 2011;6:244–285. [PubMed: 21252716]
78. Hung JJ, Yeh YC, Jeng WJ, et al. Predictive value of the international association for the study of lung cancer/American Thoracic Society/European Respiratory Society classification of lung adenocarcinoma in tumor recurrence and patient survival. *J Clin Oncol* 2014;32:2357–2364. [PubMed: 24799473]
79. Thunnissen E, Beasley MB, Borczuk A, et al. Defining Morphologic Features of Invasion in Pulmonary Nonmucinous Adenocarcinoma With Lepidic Growth: A Proposal by the International Association for the Study of Lung Cancer Pathology Committee. *J Thorac Oncol* 2023;18:447–462. [PubMed: 36503176]
80. Tsao MS, Marguet S, Le Teuff G, et al. Subtype Classification of Lung Adenocarcinoma Predicts Benefit From Adjuvant Chemotherapy in Patients Undergoing Complete Resection. *J Clin Oncol* 2015;33:3439–3446. [PubMed: 25918286]
81. Moreira AL, Ocampo PSS, Xia Y, et al. A Grading System for Invasive Pulmonary Adenocarcinoma: A Proposal From the International Association for the Study of Lung Cancer Pathology Committee. *J Thorac Oncol* 2020;15:1599–1610. [PubMed: 32562873]
82. Hayes DN, Monti S, Parmigiani G, et al. Gene expression profiling reveals reproducible human lung adenocarcinoma subtypes in multiple independent patient cohorts. *J Clin Oncol* 2006;24:5079–5090. [PubMed: 17075127]
83. Tochigi N, Dacic S, Nikiforova M, et al. Adenosquamous carcinoma of the lung: a microdissection study of KRAS and EGFR mutational and amplification status in a western patient population. *Am J Clin Pathol* 2011;135:783–789. [PubMed: 21502435]
84. Tang M, Abbas HA, Negrao MV, et al. The histologic phenotype of lung cancers is associated with transcriptomic features rather than genomic characteristics. *Nat Commun* 2021;12:7081. [PubMed: 34873156]
85. Caso R, Sanchez-Vega F, Tan KS, et al. The Underlying Tumor Genomics of Predominant Histologic Subtypes in Lung Adenocarcinoma. *J Thorac Oncol* 2020;15:1844–1856. [PubMed: 32791233]
86. Karasaki T, Moore DA, Veeriah S, et al. Evolutionary characterization of lung adenocarcinoma morphology in TRACERx. *Nat Med* 2023;29:833–845. [PubMed: 37045996]
87. Stockhammer P, Grant M, Wurtz A, et al. Co-Occurring Alterations in Multiple Tumor Suppressor Genes Are Associated With Worse Outcomes in Patients With EGFR-Mutant Lung Cancer. *J Thorac Oncol* 2024;19:240–251. [PubMed: 37806385]

88. Pros E, Saigi M, Alameda D, et al. Genome-wide profiling of non-smoking-related lung cancer cells reveals common RB1 rearrangements associated with histopathologic transformation in EGFR-mutant tumors. *Ann Oncol* 2020;31:274–282. [PubMed: 31959344]
89. Negrao MV, Araujo HA, Lamberti G, et al. Comutations and KRASG12C Inhibitor Efficacy in Advanced NSCLC. *Cancer Discov* 2023;13:1556–1571. [PubMed: 37068173]
90. Hobor S, Al Bakir M, Hiley CT, et al. Mixed responses to targeted therapy driven by chromosomal instability through p53 dysfunction and genome doubling. *Nat Commun* 2024;15:4871. [PubMed: 38871738]
91. Skoulidis F, Goldberg ME, Greenawalt DM, et al. STK11/LKB1 Mutations and PD-1 Inhibitor Resistance in KRAS-Mutant Lung Adenocarcinoma. *Cancer Discov* 2018;8:822–835. [PubMed: 29773717]
92. Foggetti G, Li C, Cai H, et al. Genetic Determinants of EGFR-Driven Lung Cancer Growth and Therapeutic Response In Vivo. *Cancer Discov* 2021;11:1736–1753. [PubMed: 33707235]
93. Trebeschi S, Drago SG, Birkbak NJ, et al. Predicting response to cancer immunotherapy using noninvasive radiomic biomarkers. *Ann Oncol* 2019;30:998–1004. [PubMed: 30895304]
94. Abbosh C, Frankell AM, Harrison T, et al. Tracking early lung cancer metastatic dissemination in TRACERx using ctDNA. *Nature* 2023;616:553–562. [PubMed: 37055640]
95. Quintanal-Villalonga A, Chan JM, Yu HA, et al. Lineage plasticity in cancer: a shared pathway of therapeutic resistance. *Nat Rev Clin Oncol* 2020;17:360–371. [PubMed: 32152485]
96. Gardner EE, Earlie EM, Li K, et al. Lineage-specific intolerance to oncogenic drivers restricts histological transformation. *Science* 2024;383:eadj1415. [PubMed: 38330136]
97. Gupta PB, Pastushenko I, Skibinski A, et al. Phenotypic Plasticity: Driver of Cancer Initiation, Progression, and Therapy Resistance. *Cell Stem Cell* 2019;24:65–78. [PubMed: 30554963]
98. Mahadevan NR, Knelson EH, Wolff JO, et al. Intrinsic Immunogenicity of Small Cell Lung Carcinoma Revealed by Its Cellular Plasticity. *Cancer Discov* 2021;11:1952–1969. [PubMed: 33707236]
99. Ishioka K, Yasuda H, Hamamoto J, et al. Upregulation of FGF9 in Lung Adenocarcinoma Transdifferentiation to Small Cell Lung Cancer. *Cancer Res* 2021;81:3916–3929. [PubMed: 34083250]
100. Mehta A, Stanger BZ. Lineage Plasticity: The New Cancer Hallmark on the Block. *Cancer Res* 2024;84:184–191. [PubMed: 37963209]
101. Park JW, Lee JK, Sheu KM, et al. Reprogramming normal human epithelial tissues to a common, lethal neuroendocrine cancer lineage. *Science* 2018;362:91–95. [PubMed: 30287662]
102. Offin M, Chan JM, Tenet M, et al. Concurrent RB1 and TP53 Alterations Define a Subset of EGFR-Mutant Lung Cancers at risk for Histologic Transformation and Inferior Clinical Outcomes. *J Thorac Oncol* 2019;14:1784–1793. [PubMed: 31228622]
103. Han X, Li F, Fang Z, et al. Transdifferentiation of lung adenocarcinoma in mice with Lkb1 deficiency to squamous cell carcinoma. *Nat Commun* 2014;5:3261. [PubMed: 24531128]
104. Tong X, Patel AS, Kim E, et al. Adeno-to-squamous transition drives resistance to KRAS inhibition in LKB1 mutant lung cancer. *Cancer Cell* 2024;42:413–428 e417. [PubMed: 38402609]
105. Malagoli Tagliazucchi G, Wiecek AJ, Withnell E, et al. Genomic and microenvironmental heterogeneity shaping epithelial-to-mesenchymal trajectories in cancer. *Nat Commun* 2023;14:789. [PubMed: 36774358]
106. Barkley D, Moncada R, Pour M, et al. Cancer cell states recur across tumor types and form specific interactions with the tumor microenvironment. *Nat Genet* 2022;54:1192–1201. [PubMed: 35931863]
107. Moghal N, Li Q, Stewart EL, et al. Single-Cell Analysis Reveals Transcriptomic Features of Drug-Tolerant Persisters and Stromal Adaptation in a Patient-Derived EGFR-Mutated Lung Adenocarcinoma Xenograft Model. *J Thorac Oncol* 2023;18:499–515. [PubMed: 36535627]
108. Hu B, Wiesehofer M, de Miguel FJ, et al. ASCL1 Drives Tolerance to Osimertinib in EGFR Mutant Lung Cancer in Permissive Cellular Contexts. *Cancer Res* 2024;84:1303–1319. [PubMed: 38359163]

109. Sharma SV, Lee DY, Li B, et al. A chromatin-mediated reversible drug-tolerant state in cancer cell subpopulations. *Cell* 2010;141:69–80. [PubMed: 20371346]
110. Guler GD, Tindell CA, Pitti R, et al. Repression of Stress-Induced LINE-1 Expression Protects Cancer Cell Subpopulations from Lethal Drug Exposure. *Cancer Cell* 2017;32:221–237 e213. [PubMed: 28781121]
111. Blakely CM, Pazarentzos E, Olivas V, et al. NF-kappaB-activating complex engaged in response to EGFR oncogene inhibition drives tumor cell survival and residual disease in lung cancer. *Cell Rep* 2015;11:98–110. [PubMed: 25843712]
112. Hellyer JA, Stehr H, Das M, et al. Impact of KEAP1/NFE2L2/CUL3 mutations on duration of response to EGFR tyrosine kinase inhibitors in EGFR mutated non-small cell lung cancer. *Lung Cancer* 2019;134:42–45. [PubMed: 31319993]
113. Kurppa KJ, Liu Y, To C, et al. Treatment-Induced Tumor Dormancy through YAP-Mediated Transcriptional Reprogramming of the Apoptotic Pathway. *Cancer Cell* 2020;37:104–122 e112. [PubMed: 31935369]
114. Maynard A, McCoach CE, Rotow JK, et al. Therapy-Induced Evolution of Human Lung Cancer Revealed by Single-Cell RNA Sequencing. *Cell* 2020;182:1232–1251 e1222. [PubMed: 32822576]
115. Binkley MS, Jeon YJ, Nesselbush M, et al. KEAP1/NFE2L2 Mutations Predict Lung Cancer Radiation Resistance That Can Be Targeted by Glutaminase Inhibition. *Cancer Discov* 2020;10:1826–1841. [PubMed: 33071215]
116. Krassikoff NE, Cowan JM, Parry DM, et al. Chromatid repulsion associated with Roberts/SC phocomelia syndrome is reduced in malignant cells and not expressed in interspecies somatic-cell hybrids. *Am J Hum Genet* 1986;39:618–630. [PubMed: 3788975]
117. Arasada RR, Shilo K, Yamada T, et al. Notch3-dependent beta-catenin signaling mediates EGFR TKI drug persistence in EGFR mutant NSCLC. *Nat Commun* 2018;9:3198. [PubMed: 30097569]
118. Suda K, Mitsudomi T. Drug Tolerance to EGFR Tyrosine Kinase Inhibitors in Lung Cancers with EGFR Mutations. *Cells* 2021;10.
119. Mikubo M, Inoue Y, Liu G, et al. Mechanism of Drug Tolerant Persister Cancer Cells: The Landscape and Clinical Implication for Therapy. *J Thorac Oncol* 2021;16:1798–1809. [PubMed: 34352380]
120. Izumi M, Costa DB, Kobayashi SS. Targeting of drug-tolerant persister cells as an approach to counter drug resistance in non-small cell lung cancer. *Lung Cancer* 2024;194:107885. [PubMed: 39002493]
121. Sasaki M, Oizumi K, Sato H, et al. [Changes in the prevalence of drug-resistant tuberculosis from 1972 to 1982]. *Kekkaku* 1985;60:241–245. [PubMed: 3928959]
122. Yang J, Antin P, Berx G, et al. Guidelines and definitions for research on epithelial-mesenchymal transition. *Nat Rev Mol Cell Biol* 2020;21:341–352. [PubMed: 32300252]
123. Song KA, Niederst MJ, Lochmann TL, et al. Epithelial-to-Mesenchymal Transition Antagonizes Response to Targeted Therapies in Lung Cancer by Suppressing BIM. *Clin Cancer Res* 2018;24:197–208. [PubMed: 29051323]
124. Tanaka K, Yu HA, Yang S, et al. Targeting Aurora B kinase prevents and overcomes resistance to EGFR inhibitors in lung cancer by enhancing BIM- and PUMA-mediated apoptosis. *Cancer Cell* 2021;39:1245–1261 e1246. [PubMed: 34388376]
125. Nilsson MB, Sun H, Robichaux J, et al. A YAP/FOXM1 axis mediates EMT-associated EGFR inhibitor resistance and increased expression of spindle assembly checkpoint components. *Sci Transl Med* 2020;12.
126. Zhang Z, Lee JC, Lin L, et al. Activation of the AXL kinase causes resistance to EGFR-targeted therapy in lung cancer. *Nat Genet* 2012;44:852–860. [PubMed: 22751098]
127. Byers LA, Diao L, Wang J, et al. An epithelial-mesenchymal transition gene signature predicts resistance to EGFR and PI3K inhibitors and identifies Axl as a therapeutic target for overcoming EGFR inhibitor resistance. *Clin Cancer Res* 2013;19:279–290. [PubMed: 23091115]
128. Konen JM, Rodriguez BL, Padhye A, et al. Dual Inhibition of MEK and AXL Targets Tumor Cell Heterogeneity and Prevents Resistant Outgrowth Mediated by the Epithelial-to-Mesenchymal Transition in NSCLC. *Cancer Res* 2021;81:1398–1412. [PubMed: 33402388]

129. Kitai H, Ebi H, Tomida S, et al. Epithelial-to-Mesenchymal Transition Defines Feedback Activation of Receptor Tyrosine Kinase Signaling Induced by MEK Inhibition in KRAS-Mutant Lung Cancer. *Cancer Discov* 2016;6:754–769. [PubMed: 27154822]
130. Raoof S, Mulford IJ, Frisco-Cabanas H, et al. Targeting FGFR overcomes EMT-mediated resistance in EGFR mutant non-small cell lung cancer. *Oncogene* 2019;38:6399–6413. [PubMed: 31324888]
131. Recondo G, Mezquita L, Facchinetti F, et al. Diverse Resistance Mechanisms to the Third-Generation ALK Inhibitor Lorlatinib in ALK-Rearranged Lung Cancer. *Clin Cancer Res* 2020;26:242–255. [PubMed: 31585938]
132. Ortiz-Cuaran S, Swalduz A, Foy JP, et al. Epithelial-to-mesenchymal transition promotes immune escape by inducing CD70 in non-small cell lung cancer. *Eur J Cancer* 2022;169:106–122. [PubMed: 35550950]
133. Nilsson MB, Yang Y, Heeke S, et al. CD70 is a therapeutic target upregulated in EMT-associated EGFR tyrosine kinase inhibitor resistance. *Cancer Cell* 2023;41:340–355 e346. [PubMed: 36787696]
134. Federico L, McGrail DJ, Bentebibel SE, et al. Distinct tumor-infiltrating lymphocyte landscapes are associated with clinical outcomes in localized non-small-cell lung cancer. *Ann Oncol* 2022;33:42–56. [PubMed: 34653632]
135. Enfield KSS, Colliver E, Lee C, et al. Spatial Architecture of Myeloid and T Cells Orchestrates Immune Evasion and Clinical Outcome in Lung Cancer. *Cancer Discov* 2024;14:1018–1047. [PubMed: 38581685]
136. Rizvi NA, Hellmann MD, Snyder A, et al. Cancer immunology. Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. *Science* 2015;348:124–128. [PubMed: 25765070]
137. Thommen DS, Koelzer VH, Herzig P, et al. A transcriptionally and functionally distinct PD-1(+) CD8(+) T cell pool with predictive potential in non-small-cell lung cancer treated with PD-1 blockade. *Nat Med* 2018;24:994–1004. [PubMed: 29892065]
138. Anagnostou V, Smith KN, Forde PM, et al. Evolution of Neoantigen Landscape during Immune Checkpoint Blockade in Non-Small Cell Lung Cancer. *Cancer Discov* 2017;7:264–276. [PubMed: 28031159]
139. McGranahan N, Rosenthal R, Hiley CT, et al. Allele-Specific HLA Loss and Immune Escape in Lung Cancer Evolution. *Cell* 2017;171:1259–1271 e1211. [PubMed: 29107330]
140. Chen HL, Gabrilovich D, Tampe R, et al. A functionally defective allele of TAP1 results in loss of MHC class I antigen presentation in a human lung cancer. *Nat Genet* 1996;13:210–213. [PubMed: 8640228]
141. Rosenthal R, Cadieux EL, Salgado R, et al. Neoantigen-directed immune escape in lung cancer evolution. *Nature* 2019;567:479–485. [PubMed: 30894752]
142. Pennycuik A, Teixeira VH, AbdulJabbar K, et al. Immune Surveillance in Clinical Regression of Preinvasive Squamous Cell Lung Cancer. *Cancer Discov* 2020;10:1489–1499. [PubMed: 32690541]
143. AbdulJabbar K, Raza SEA, Rosenthal R, et al. Geospatial immune variability illuminates differential evolution of lung adenocarcinoma. *Nat Med* 2020;26:1054–1062. [PubMed: 32461698]
144. Leader AM, Grout JA, Maier BB, et al. Single-cell analysis of human non-small cell lung cancer lesions refines tumor classification and patient stratification. *Cancer Cell* 2021;39:1594–1609 e1512. [PubMed: 34767762]
145. Grout JA, Sirven P, Leader AM, et al. Spatial Positioning and Matrix Programs of Cancer-Associated Fibroblasts Promote T-cell Exclusion in Human Lung Tumors. *Cancer Discov* 2022;12:2606–2625. [PubMed: 36027053]
146. Saigi M, Mate JL, Carcereny E, et al. HLA-I levels correlate with survival outcomes in response to immune checkpoint inhibitors in non-small cell lung cancer. *Lung Cancer* 2024;189:107502. [PubMed: 38359742]

147. Pereira C, Gimenez-Xavier P, Pros E, et al. Genomic Profiling of Patient-Derived Xenografts for Lung Cancer Identifies B2M Inactivation Impairing Immunorecognition. *Clin Cancer Res* 2017;23:3203–3213. [PubMed: 28302866]
148. Cords L, Engler S, Haberecker M, et al. Cancer-associated fibroblast phenotypes are associated with patient outcome in non-small cell lung cancer. *Cancer Cell* 2024;42:396–412 e395. [PubMed: 38242124]
149. Simoni Y, Becht E, Fehlings M, et al. Bystander CD8(+) T cells are abundant and phenotypically distinct in human tumour infiltrates. *Nature* 2018;557:575–579. [PubMed: 29769722]
150. Caushi JX, Zhang J, Ji Z, et al. Author Correction: Transcriptional programs of neoantigen-specific TIL in anti-PD-1-treated lung cancers. *Nature* 2021;598:E1. [PubMed: 34608287]
151. Scheper W, Kelderman S, Fanchi LF, et al. Low and variable tumor reactivity of the intratumoral TCR repertoire in human cancers. *Nat Med* 2019;25:89–94. [PubMed: 30510250]
152. van Weverwijk A, de Visser KE. Mechanisms driving the immunoregulatory function of cancer cells. *Nat Rev Cancer* 2023;23:193–215. [PubMed: 36717668]
153. Albuquerque-Bejar JJ, Navajas-Chocarro P, Saigi M, et al. MYC activation impairs cell-intrinsic IFN γ signaling and confers resistance to anti-PD1/PD-L1 therapy in lung cancer. *Cell Rep Med* 2023;4:101006. [PubMed: 37044092]
154. Saigi M, Albuquerque-Bejar JJ, Mc Leer-Florin A, et al. MET-Oncogenic and JAK2-Inactivating Alterations Are Independent Factors That Affect Regulation of PD-L1 Expression in Lung Cancer. *Clin Cancer Res* 2018;24:4579–4587. [PubMed: 29898990]
155. Dammeijer F, van Gulijk M, Mulder EE, et al. The PD-1/PD-L1-Checkpoint Restrains T cell Immunity in Tumor-Draining Lymph Nodes. *Cancer Cell* 2020;38:685–700 e688. [PubMed: 33007259]
156. Horton BL, Morgan DM, Momin N, et al. Lack of CD8(+) T cell effector differentiation during priming mediates checkpoint blockade resistance in non-small cell lung cancer. *Sci Immunol* 2021;6:eabi8800. [PubMed: 34714687]
157. Liu B, Hu X, Feng K, et al. Temporal single-cell tracing reveals clonal revival and expansion of precursor exhausted T cells during anti-PD-1 therapy in lung cancer. *Nat Cancer* 2022;3:108–121. [PubMed: 35121991]
158. Pai JA, Hellmann MD, Sauter JL, et al. Lineage tracing reveals clonal progenitors and long-term persistence of tumor-specific T cells during immune checkpoint blockade. *Cancer Cell* 2023;41:776–790 e777. [PubMed: 37001526]
159. Wu TD, Madireddi S, de Almeida PE, et al. Peripheral T cell expansion predicts tumour infiltration and clinical response. *Nature* 2020;579:274–278. [PubMed: 32103181]
160. Zagorulya M, Yim L, Morgan DM, et al. Tissue-specific abundance of interferon-gamma drives regulatory T cells to restrain DC1-mediated priming of cytotoxic T cells against lung cancer. *Immunity* 2023;56:386–405 e310. [PubMed: 36736322]
161. van Gulijk M, van Krimpen A, Schetters S, et al. PD-L1 checkpoint blockade promotes regulatory T cell activity that underlies therapy resistance. *Sci Immunol* 2023;8:eabn6173. [PubMed: 37205768]
162. Maier B, Leader AM, Chen ST, et al. A conserved dendritic-cell regulatory program limits antitumour immunity. *Nature* 2020;580:257–262. [PubMed: 32269339]
163. Teillaud JL, Houel A, Panouillot M, et al. Tertiary lymphoid structures in anticancer immunity. *Nat Rev Cancer* 2024;24:629–646. [PubMed: 39117919]
164. Chen JH, Nieman LT, Spurrell M, et al. Human lung cancer harbors spatially organized stem-immunity hubs associated with response to immunotherapy. *Nat Immunol* 2024;25:644–658. [PubMed: 38503922]
165. Memon D, Schoenfeld AJ, Ye D, et al. Clinical and molecular features of acquired resistance to immunotherapy in non-small cell lung cancer. *Cancer Cell* 2024;42:209–224 e209. [PubMed: 38215748]
166. Gettinger S, Choi J, Hastings K, et al. Impaired HLA Class I Antigen Processing and Presentation as a Mechanism of Acquired Resistance to Immune Checkpoint Inhibitors in Lung Cancer. *Cancer Discov* 2017;7:1420–1435. [PubMed: 29025772]

167. Skoulidis F, Araujo HA, Do MT, et al. CTLA4 blockade abrogates KEAP1/STK11-related resistance to PD-(L)1 inhibitors. *Nature* 2024;635:462–471. [PubMed: 39385035]
168. Hiltbrunner S, Cords L, Kasser S, et al. Acquired resistance to anti-PD1 therapy in patients with NSCLC associates with immunosuppressive T cell phenotype. *Nat Commun* 2023;14:5154. [PubMed: 37620318]
169. Mathew D, Marmarelis ME, Foley C, et al. Combined JAK inhibition and PD-1 immunotherapy for non-small cell lung cancer patients. *Science* 2024;384:eadf1329. [PubMed: 38900877]
170. Sumii M, Namba M, Tokumo K, et al. Concurrent Mutations in STK11 and KEAP1 Cause Treatment Resistance in KRAS Wild-type Non-small-cell Lung Cancer. *Intern Med* 2023;62:3001–3004. [PubMed: 36858519]
171. Wohlieter CA, Richards AL, Uddin F, et al. Concurrent Mutations in STK11 and KEAP1 Promote Ferroptosis Protection and SCD1 Dependence in Lung Cancer. *Cell Rep* 2020;33:108444. [PubMed: 33264619]
172. Combs JA, DeNicola GM. The Non-Essential Amino Acid Cysteine Becomes Essential for Tumor Proliferation and Survival. *Cancers (Basel)* 2019;11.
173. Greenwood HE, Barber AR, Edwards RS, et al. Imaging NRF2 activation in non-small cell lung cancer with positron emission tomography. *Nat Commun* 2024;15:10484. [PubMed: 39690148]
174. Pons-Tostivint E, Lugat A, Fontenau JF, et al. STK11/LKB1 Modulation of the Immune Response in Lung Cancer: From Biology to Therapeutic Impact. *Cells* 2021;10.
175. Hu L, Liu M, Tang B, et al. Posttranslational regulation of liver kinase B1 in human cancer. *J Biol Chem* 2023;299:104570. [PubMed: 36870679]
176. Hollstein PE, Eichner LJ, Brun SN, et al. The AMPK-Related Kinases SIK1 and SIK3 Mediate Key Tumor-Suppressive Effects of LKB1 in NSCLC. *Cancer Discov* 2019;9:1606–1627. [PubMed: 31350328]
177. Sun Z, Jiang Q, Li J, et al. The potent roles of salt-inducible kinases (SIKs) in metabolic homeostasis and tumorigenesis. *Signal Transduct Target Ther* 2020;5:150. [PubMed: 32788639]
178. Pillai R, Hayashi M, Zavitsanou AM, et al. NRF2: KEAPing Tumors Protected. *Cancer Discov* 2022;12:625–643. [PubMed: 35101864]
179. Sen U, Coleman C, Sen T. Stearoyl coenzyme A desaturase-1: multitasker in cancer, metabolism, and ferroptosis. *Trends Cancer* 2023;9:480–489. [PubMed: 37029018]
180. Lee HM, Muhammad N, Lieu EL, et al. Concurrent loss of LKB1 and KEAP1 enhances SHMT-mediated antioxidant defence in KRAS-mutant lung cancer. *Nat Metab* 2024;6:1310–1328. [PubMed: 38877143]
181. Yu T, Liu Z, Tao Q, et al. Targeting tumor-intrinsic SLC16A3 to enhance anti-PD-1 efficacy via tumor immune microenvironment reprogramming. *Cancer Lett* 2024;589:216824. [PubMed: 38522774]
182. Yoshida GJ. Metabolic reprogramming: the emerging concept and associated therapeutic strategies. *J Exp Clin Cancer Res* 2015;34:111. [PubMed: 26445347]
183. Harmon C, Robinson MW, Hand F, et al. Lactate-Mediated Acidification of Tumor Microenvironment Induces Apoptosis of Liver-Resident NK Cells in Colorectal Liver Metastasis. *Cancer Immunol Res* 2019;7:335–346. [PubMed: 30563827]
184. Kumar A, Pyaram K, Yarosz EL, et al. Enhanced oxidative phosphorylation in NKT cells is essential for their survival and function. *Proc Natl Acad Sci U S A* 2019;116:7439–7448. [PubMed: 30910955]
185. Nasi A, Fekete T, Krishnamurthy A, et al. Dendritic cell reprogramming by endogenously produced lactic acid. *J Immunol* 2013;191:3090–3099. [PubMed: 23956421]
186. Morrot A, da Fonseca LM, Salustiano EJ, et al. Metabolic Symbiosis and Immunomodulation: How Tumor Cell-Derived Lactate May Disturb Innate and Adaptive Immune Responses. *Front Oncol* 2018;8:81. [PubMed: 29629338]
187. Liao ZX, Kempson IM, Hsieh CC, et al. Potential therapeutics using tumor-secreted lactate in nonsmall cell lung cancer. *Drug Discov Today* 2021;26:2508–2514. [PubMed: 34325010]
188. Schwickert G, Walenta S, Sundfor K, et al. Correlation of high lactate levels in human cervical cancer with incidence of metastasis. *Cancer Res* 1995;55:4757–4759. [PubMed: 7585499]

189. Hur H, Xuan Y, Kim YB, et al. Expression of pyruvate dehydrogenase kinase-1 in gastric cancer as a potential therapeutic target. *Int J Oncol* 2013;42:44–54. [PubMed: 23135628]
190. Chang YC, Chan MH, Yang YF, et al. Glucose transporter 4: Insulin response mastermind, glycolysis catalyst and treatment direction for cancer progression. *Cancer Lett* 2023;563:216179. [PubMed: 37061122]
191. Spencer TL, Lehninger AL. L-lactate transport in Ehrlich ascites-tumour cells. *Biochem J* 1976;154:405–414. [PubMed: 7237]
192. Bosshart PD, Kalbermatter D, Bonetti S, et al. Mechanistic basis of L-lactate transport in the SLC16 solute carrier family. *Nat Commun* 2019;10:2649. [PubMed: 31201333]
193. Markou A, Tzanikou E, Kallergi G, et al. Evaluation of Monocarboxylate Transporter 4 (MCT4) Expression and Its Prognostic Significance in Circulating Tumor Cells From Patients With Early Stage Non-Small-Cell Lung Cancer. *Front Cell Dev Biol* 2021;9:641978. [PubMed: 33968927]
194. Doherty JR, Yang C, Scott KE, et al. Blocking lactate export by inhibiting the Myc target MCT1 Disables glycolysis and glutathione synthesis. *Cancer Res* 2014;74:908–920. [PubMed: 24285728]
195. Polanski R, Hodgkinson CL, Fusi A, et al. Activity of the monocarboxylate transporter 1 inhibitor AZD3965 in small cell lung cancer. *Clin Cancer Res* 2014;20:926–937. [PubMed: 24277449]
196. Jiang J, Huang D, Jiang Y, et al. Lactate Modulates Cellular Metabolism Through Histone Lactylation-Mediated Gene Expression in Non-Small Cell Lung Cancer. *Front Oncol* 2021;11:647559. [PubMed: 34150616]
197. Chen D, Zhang X, Li Z, et al. Metabolic regulatory crosstalk between tumor microenvironment and tumor-associated macrophages. *Theranostics* 2021;11:1016–1030. [PubMed: 33391518]
198. Wang L, Ma Y, Zhang S, et al. NFATc2 promotes lactate and M2 macrophage polarization through USP17 in lung adenocarcinoma. *Anticancer Drugs* 2024;35:385–396. [PubMed: 38386130]
199. Dowling CM, Zhang H, Chonghaile TN, et al. Shining a light on metabolic vulnerabilities in non-small cell lung cancer. *Biochim Biophys Acta Rev Cancer* 2021;1875:188462. [PubMed: 33130228]
200. Kim JH, Nam B, Choi YJ, et al. Enhanced Glycolysis Supports Cell Survival in EGFR-Mutant Lung Adenocarcinoma by Inhibiting Autophagy-Mediated EGFR Degradation. *Cancer Res* 2018;78:4482–4496. [PubMed: 29945964]
201. Fernandes ET, Hollabaugh RS, Boulden T. Mediastinal mass and radiolucent esophageal foreign body. *J Pediatr Surg* 1989;24:1135–1136. [PubMed: 2809984]
202. Apicella M, Giannoni E, Fiore S, et al. Increased Lactate Secretion by Cancer Cells Sustains Non-cell-autonomous Adaptive Resistance to MET and EGFR Targeted Therapies. *Cell Metab* 2018;28:848–865 e846. [PubMed: 30174307]
203. Zhang J, Song F, Zhao X, et al. EGFR modulates monounsaturated fatty acid synthesis through phosphorylation of SCD1 in lung cancer. *Mol Cancer* 2017;16:127. [PubMed: 28724430]
204. Ma Y, Yu C, Mohamed EM, et al. A causal link from ALK to hexokinase II overexpression and hyperactive glycolysis in EML4-ALK-positive lung cancer. *Oncogene* 2016;35:6132–6142. [PubMed: 27132509]
205. Strohecker AM, White E. Autophagy promotes BrafV600E-driven lung tumorigenesis by preserving mitochondrial metabolism. *Autophagy* 2014;10:384–385. [PubMed: 24362353]
206. Strohecker AM, Guo JY, Karsli-Uzunbas G, et al. Autophagy sustains mitochondrial glutamine metabolism and growth of BrafV600E-driven lung tumors. *Cancer Discov* 2013;3:1272–1285. [PubMed: 23965987]
207. Scheiermann J, Klinman DM. Three distinct pneumotypes characterize the microbiome of the lung in BALB/cJ mice. *PLoS One* 2017;12:e0180561. [PubMed: 28683098]
208. Pezzulo AA, Kelly PH, Nassar BS, et al. Abundant DNase I-sensitive bacterial DNA in healthy porcine lungs and its implications for the lung microbiome. *Appl Environ Microbiol* 2013;79:5936–5941. [PubMed: 23872563]
209. Gollwitzer ES, Saglani S, Trompette A, et al. Lung microbiota promotes tolerance to allergens in neonates via PD-L1. *Nat Med* 2014;20:642–647. [PubMed: 24813249]

210. Pattaroni C, Watzenboeck ML, Schneidegger S, et al. Early-Life Formation of the Microbial and Immunological Environment of the Human Airways. *Cell Host Microbe* 2018;24:857–865 e854. [PubMed: 30503510]
211. Nagasaka M, Sexton R, Alhasan R, et al. Gut microbiome and response to checkpoint inhibitors in non-small cell lung cancer-A review. *Crit Rev Oncol Hematol* 2020;145:102841. [PubMed: 31884204]
212. Khan FH, Bhat BA, Sheikh BA, et al. Microbiome dysbiosis and epigenetic modulations in lung cancer: From pathogenesis to therapy. *Semin Cancer Biol* 2022;86:732–742. [PubMed: 34273520]
213. Boursi B, Mamtani R, Haynes K, et al. Recurrent antibiotic exposure may promote cancer formation--Another step in understanding the role of the human microbiota? *Eur J Cancer* 2015;51:2655–2664. [PubMed: 26338196]
214. Elkrief A, Mendez-Salazar EO, Maillou J, et al. Antibiotics are associated with worse outcomes in lung cancer patients treated with chemotherapy and immunotherapy. *NPJ Precis Oncol* 2024;8:143. [PubMed: 39014160]
215. Shiels MS, Albanes D, Virtamo J, et al. Increased risk of lung cancer in men with tuberculosis in the alpha-tocopherol, beta-carotene cancer prevention study. *Cancer Epidemiol Biomarkers Prev* 2011;20:672–678. [PubMed: 21335509]
216. Liu HX, Tao LL, Zhang J, et al. Difference of lower airway microbiome in bilateral protected specimen brush between lung cancer patients with unilateral lobar masses and control subjects. *Int J Cancer* 2018;142:769–778. [PubMed: 29023689]
217. Yu G, Gail MH, Consonni D, et al. Characterizing human lung tissue microbiota and its relationship to epidemiological and clinical features. *Genome Biol* 2016;17:163. [PubMed: 27468850]
218. Ma QY, Huang DY, Zhang HJ, et al. Upregulation of bacterial-specific Th1 and Th17 responses that are enriched in CXCR5(+)CD4(+) T cells in non-small cell lung cancer. *Int Immunopharmacol* 2017;52:305–309. [PubMed: 28982050]
219. Routy B, Le Chatelier E, Derosa L, et al. Gut microbiome influences efficacy of PD-1-based immunotherapy against epithelial tumors. *Science* 2018;359:91–97. [PubMed: 29097494]
220. Gunjur A, Shao Y, Rozday T, et al. A gut microbial signature for combination immune checkpoint blockade across cancer types. *Nat Med* 2024;30:797–809. [PubMed: 38429524]

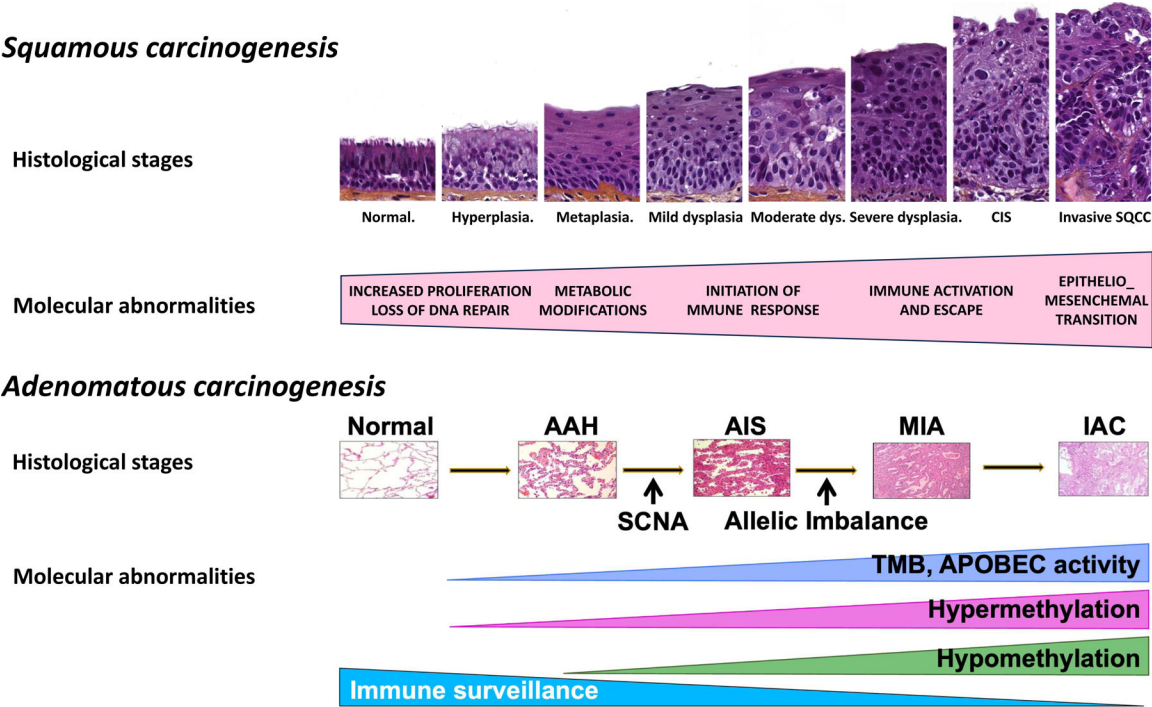


Figure 1. Multi-stage lung carcinogenesis. The chronological histological changes observed in lung squamous and adenomatous carcinogenesis and their cumulative molecular changes. Abbreviations: Dys, dysplasia; CIS, carcinoma in situ; SQCC, squamous cell carcinoma; TMB: tumor mutation burden; SCNA: somatic copy number alteration; AAH: atypical adenomatous hyperplasia; AIS: adenocarcinoma in situ; MIA: minimally invasive adenocarcinoma; IAC invasive adenocarcinoma. Hypomethylation and Hypermethylation: these refer to aberrant methylation changes compared to matched normal lung tissues.

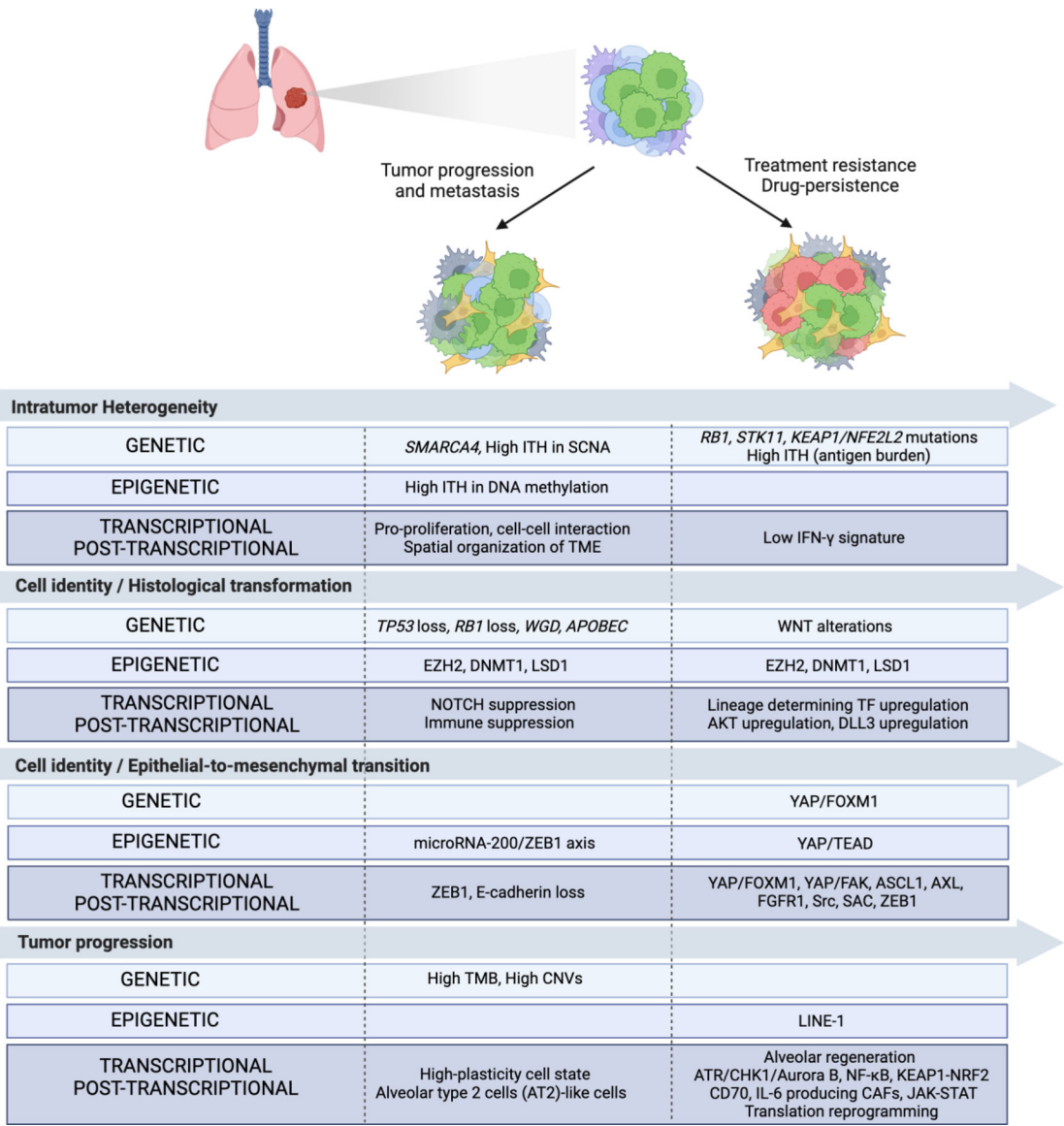


Figure 2. Molecular regulators and potential therapeutic targets of tumor heterogeneity and plasticity in lung cancer. Graphical representation of genetic, epigenetic, transcriptional and post-transcriptional mechanisms that promote tumor progression, metastasis and resistance to therapy. Abbreviations: SCNA: Somatic copy number alterations; ITH: Intratumor heterogeneity; TME: Tumor microenvironment; CNV: copy number variations; TMB: tumor mutational burden; SAC: Spindle assembly checkpoint proteins; CAFs: Cancer-associated fibroblasts.

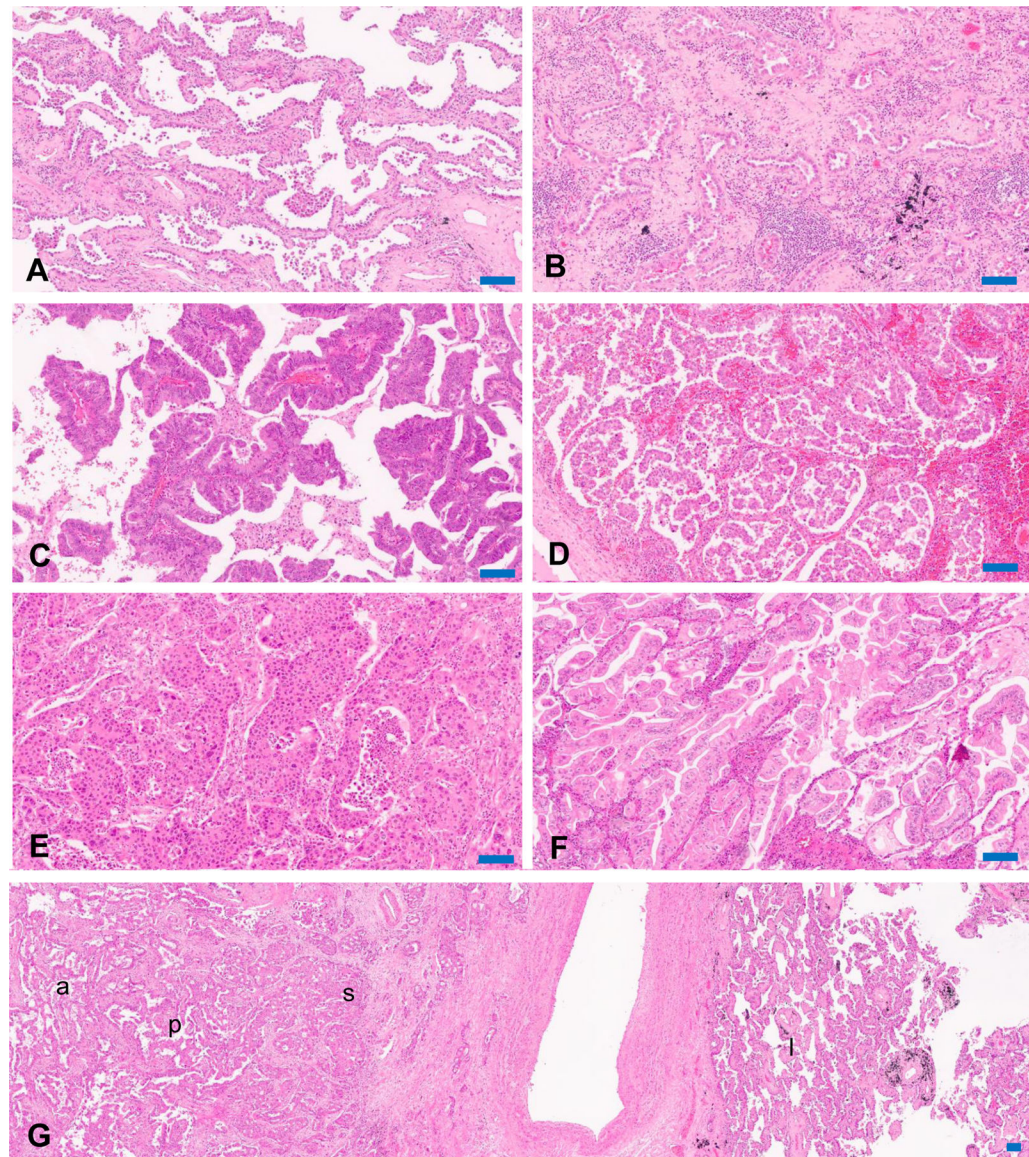


Figure 3.

Heterogeneous growth patterns observed in LUADs. (A) Lepidic pattern is considered in-situ non-invasive growth, while (B) acinar, (C) papillary, (D) micropapillary and (E) solid patterns are regarded as invasive growth patterns. (F) Mucinous adenocarcinoma is distinguished for its finely vacuolated cytoplasm that contains mucin substance. (G) Up to 80% of non-mucinous LUAD contains a mixture of multiple patterns. The predominant pattern is used to subtype non-mucinous LUAD, and their relative abundance is used for grading the tumor. Abbreviations: lep: lepidic; acn: acinar; pap: papillary; sol: solid. Scale bar: 100 μ m

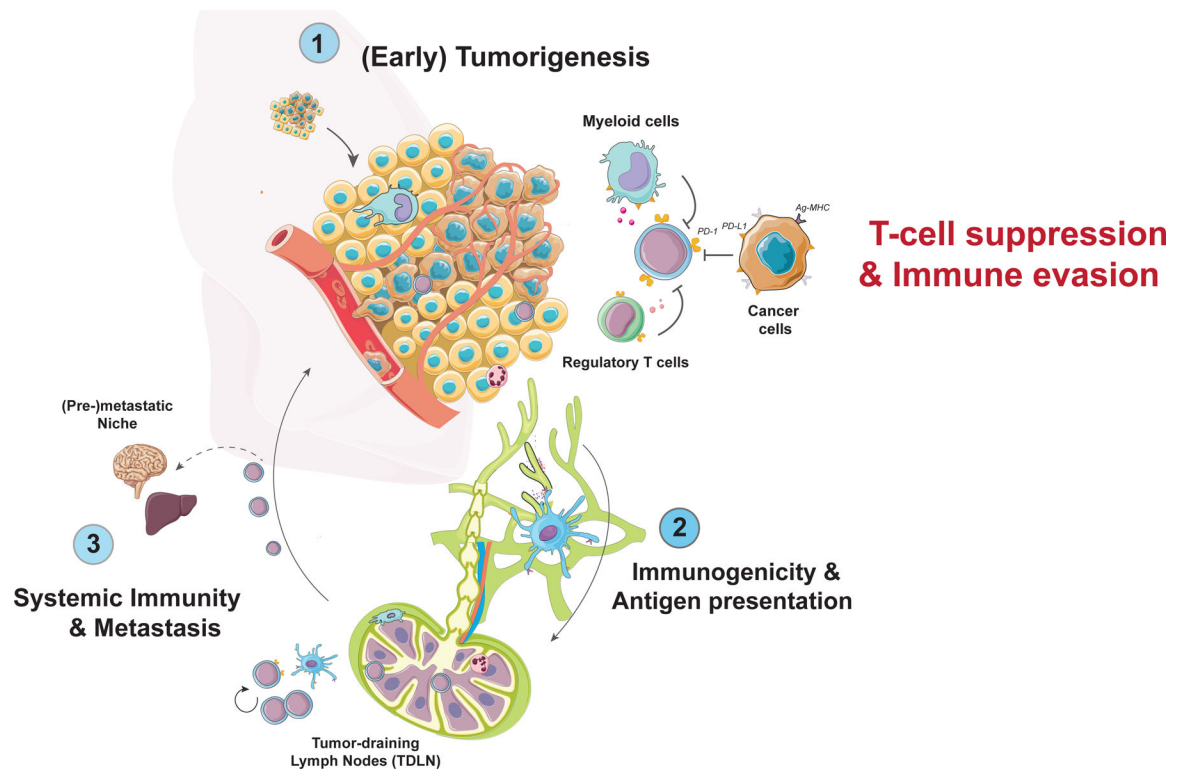


Figure 4.

Immune evasion as a hallmark of lung cancer progression. (1) Evasion of anti-tumor immunity occurs early during tumor progression in preneoplastic lesions through a plethora of different mechanisms including contact-dependent [e.g. PD-1/PD-L1 checkpoint interaction, loss of immunogenicity through MHC- or neo-antigen (Ag) loss)] or contact-independent processes (e.g. suppressive cytokines, metabolites etc.). (2) These mechanisms extend beyond the tumor microenvironment (TME) and affect antigen presentation in tumor-draining lymph nodes (TDLNs) and systemic anti-tumor immunity. (3) Furthermore, several immune cells either positively (e.g. circulating tumor-specific CD8⁺ T cells) or negatively (e.g. neutrophils) contribute to distant organ metastases through conditioning of the (pre) metastatic niche.

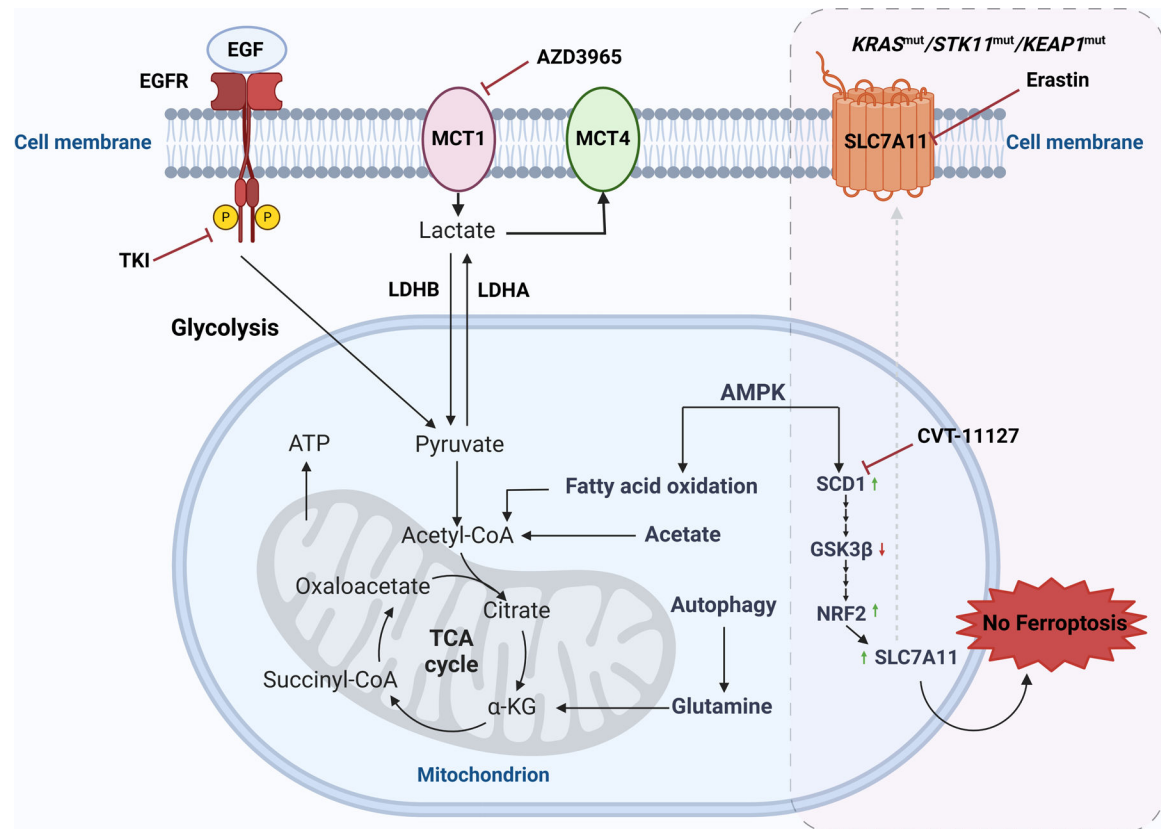


Figure 5.

Metabolic reprogramming in EGFR and KRAS driven NSCLC. EGFR-mutant NSCLC demonstrates a high dependency on glucose, glutamine and lactate metabolism, in addition to fatty acid synthesis. Monocarboxylate transporters are lactic acid transporters, which mediate the influx and efflux of lactate in tumor cells. MCT1 inhibitors (e.g. AZD3965) have preclinical activity in lung cancer cells by inhibition of the immunosuppressive effects driven by lactate metabolism. The interplay between STK11 and KEAP1 co-mutations in KRAS-mutant NSCLC can lead to rewiring of cellular metabolism and redox homeostasis dysregulation, influencing cancer progression and treatment response. Ferroptosis evasion is enriched in STK11 and KEAP1 co-mutated lung cancer cells, where several genes regulated by NRF2 encode proteins implicated in ferroptosis control, including SLC7A11, a key regulator of glutathione metabolism. Targeted inhibition of SCD1 activity by CMT-11127 can disrupt ferroptosis in STK11 and KEAP1 co-mutated cells. Further exploration of NSCLC metabolic plasticity may enable the strategic use of novel metabolic inhibitors, alone or in combination with immunotherapies, to overcome treatment resistance and enhance therapeutic efficacy.