

Model of Induced Leakage of Polymethylmethacrylate Inside Epidural Space and Prevertebral Muscles During Vertebroplasty in Pigs: Clinical, Macroscopical, and Histological Study

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Study Design: Experimental study in animals.

Purpose: Study the clinical behavior of animals after an induced leakage of cement during vertebroplasty in pigs. Study the distribution of polymethylmethacrylate inside the epidural space and prevertebral muscle. Study the histological findings of the spinal cord and muscles, which contact with cement.

Overview of Literature: Although vertebroplasty has a low rate of complication, leakage of cement is highly frequent. There is paucity, in how cement is distributed inside the spinal canal and what occurs when soft tissue comes into contact with polymethylmethacrylate.

Methods: We performed vertebroplasty on six pigs. We performed a leakage of cement into the epidural space and into prevertebral muscles. Two weeks later we performed an anatomic evaluation regarding the spreading of polymethylmethacrylate and a histological analysis of soft tissues that came into contact with it.

Results: No clinical alterations were observed. We observed a laminar distribution of the cement surrounding dura mater, and creating a fusiform cavity inside muscles. Spinal cord was normal in all the animals. In dura mater, we observed: synovial metaplasia, inflammatory reaction, crystal deposits, and giant-cell-reaction. In muscles, we observed: inflammatory reaction, crystal deposits, giant-cell-reaction, muscular atrophy, fibrosis, and synovial metaplasia.

Conclusions: The spinal cord was normal; it is likely that dura mater and cerebrospinal fluid are responsible to isolate neural structures from cement. Dura mater and muscle showed similar histological changes than other publications. Synovial metaplasia was observed in dura mater and muscles that came into contact with cement. The pulsatile rubbing between the tissue and cement could be responsible of this phenomenon.

Keywords: Spinal cord; Psoas muscles; Histology; Vertebroplasty; Polymethylmethacrylate

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Introduction

Vertebroplasty is a useful technique for treating osteoporotic compression fractures, vertebral malignancy, and symptomatic vertebral hemangiomas. Excellent rates of pain relief and improved quality of life have been cited throughout various studies [1-7].

Although vertebroplasty have a low rate of complications, 1% to 3% for osteoporotic fractures, 2.5% for hemangiomas, and up to 10% for metastatic vertebral lesions [8-12], leakage of cement out of the bone towards the epidural space, vessels, or the perivertebral tissues is highly frequent. It has been observed in up to 70% of the cases, fortunately without severe clinical complications [11-16].

Existing literature poorly describes the behavior of the cement when it leaks from the vertebrae and also poorly describes the histological changes of the soft tissue like dura mater and muscle when they come into contact with polymethylmethacrylate. Most of the studies, in humans and in some animals, have focused on bony histological changes, describing inflammatory reaction, foreign body reaction and bone necrosis [17-23].

The high frequency of leakage of polymethylmethacrylate during vertebroplasty and the paucity of information especially on the macroscopic distribution of the cement in the epidural space and the potential histological alterations of the spinal soft tissues, like the spinal cord, the dura mater and the perivertebral muscles, that may come into contact with the cement when it leaks. Considering this, we decided to set up a reproducible model of epidural and prevertebral leakage of polymethylmethacrylate for vertebroplasty in pigs, to study the cement-spreading pattern into the spinal canal and muscles, the potential histological changes in the spinal soft tissues and try to relate it, with the clinical behavior of the animals after the procedure.

Materials and Methods

For this study we obtained approval from the local institutional review board for animal experiments.

1. Animals

We used six female, three-month-old Large White Landrace pigs weighing 30 to 35 kg, obtained from a professional stockbreeder.

We followed the rules for assessment and alleviation of postoperative pain and end point or euthanasia criteria for pain behavior, paraplegia, and dying animal models [24,25].

The surgical procedure began with a preanesthetic sedation (ketamine 10 mg/kg, intramuscular one time) administered in the stall, followed by general anesthesia that was administered by orotracheal intubation as follows: anesthetic induction: azaperone 2 mg/kg intramuscular one time, atropine 1 mg intramuscular one time, etomidate 3 mg/kg intravenous, then, muscle relaxation with pancuronium 0.2 mg/kg intravenously, followed by orotracheal intubation.

The maintenance of the anesthesia was done with isoflurane 0.8% to 1% plus oxygen 40% and pancuronium 0.2 mg/kg every 30 minutes intravenously plus fentanyl 0.005 mg/kg intravenously every 30 minutes.

Like prophylactic antibiotics we used one dose of 1 g intravenous ampicillin.

For the surgical procedure the animals were placed on a radiolucent surgical table in a prone position.

The animal's back was shaved, disinfected with chlorhexidine, and draped with surgical fields, exposing the dorsal mid line.

We used C-arm radioscopic guidance (Powermobil Siemens, Erlangen, Germany). For vertebroplasty we used a conventional 15 cm, 13-gage vertebroplasty trocars (Osteo-Site, Cook, IN, USA), vertebroplasty cement (Vertebroplastic, De Puy Spine, Raynham, MA, USA) and 2 mL Luer-locked syringes for the cement injection. The cement was mixed and injected following the manufacturer's instructions.

To select the vertebral level we choose the vertebra most easily viewed under antero-posterior C-arm imaging, currently between L1 and L3.

To place the cement in contact with prevertebral and myelofurcular tissues, we performed two types of cement escape: one was an epidural leak, for this, we performed a percutaneous right pedicle cannulation of a central lumbar vertebra, later on, under the antero-posterior radiological view, the tip of the trocar penetrates the pedicle from the lateral to the medial cortex following a 30°-angle trajectory, a lateral view was used to corroborate the epidural placement of the tip (Fig. 1).

To induce a prevertebral leak, we performed a percutaneous access on the left pedicle of an adjacent vertebra. Under the antero-posterior radiological view the trocar

penetrates the center of the pedicle in a near 90°-angle trajectory, later on, using a lateral radiological view, we penetrated the anterior cortex of the vertebra (Fig. 1).

Under antero-posterior and lateral radiological views, one milliliter of vertebroplasty cement was injected in both vertebrae.

Finally the skin was disinfected with clorhexidine and sutured with surgical silk 3/0.

After surgery the animals were placed in a warm cage and received postoperative analgesia (buprenorphine

0,005 mg/kg subcutaneous, one time). The animals were placed in their cages with free food and water delivery, and were observed daily for limb motor function using the Tarlov scales [26], sphincters function, and any pain behavior or end point criteria [24,25].

2. Euthanasia

The animals were sacrificed two weeks after the surgery with an intravenous injection of pentobarbital sodium

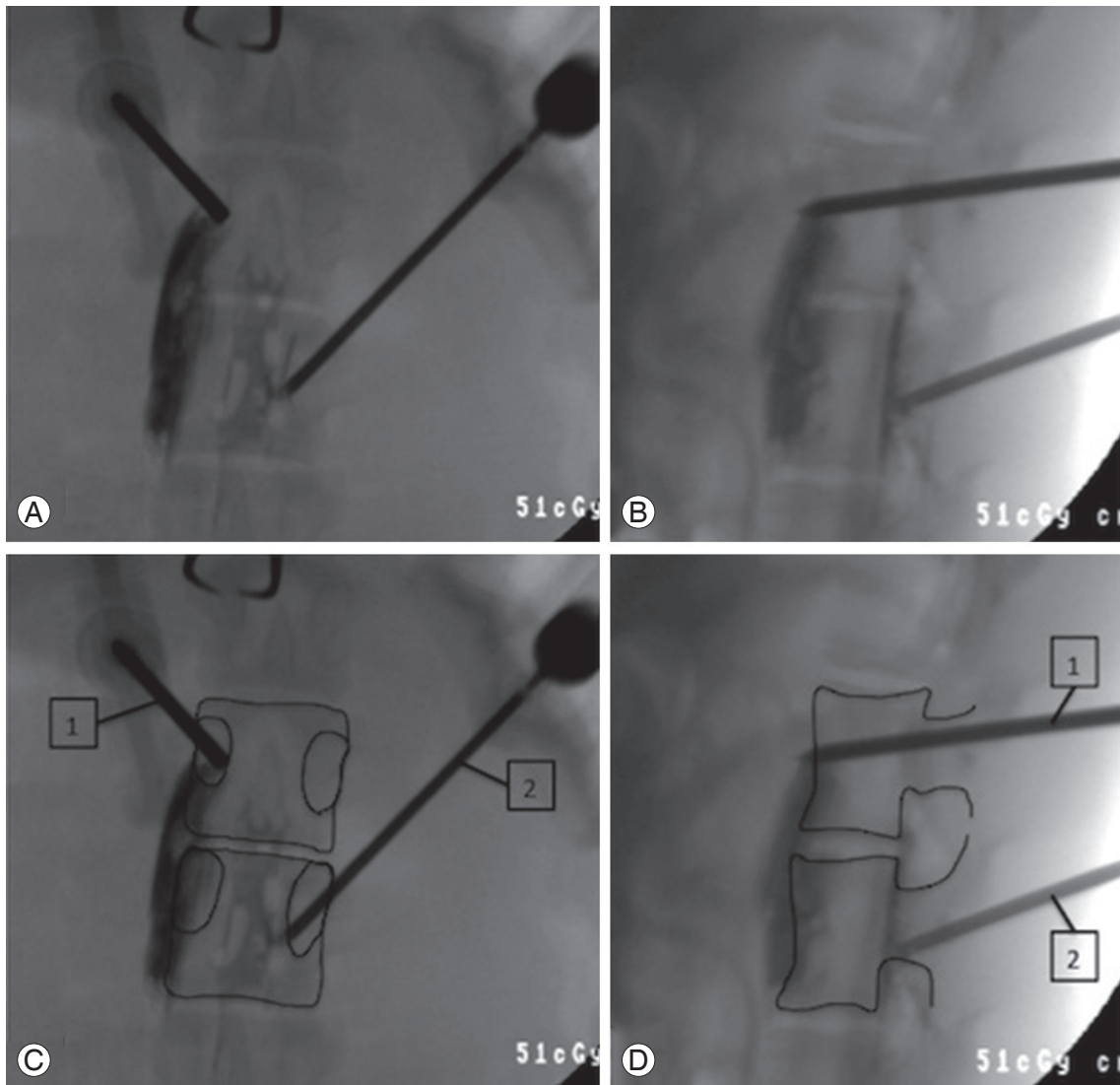


Fig. 1. (A, B) Intraoperative X-ray. (C, D) Diagrams enhancing the vertebral cortex. (A, C) Frontal view, upper vertebra, trocar number 1 (prevertebral leak) the trocar enters the pedicle in a near right angle trajectory. Inferior vertebrae, trocar number 2 (epidural leak): the cannula enters the pedicle in a 30-degree inward angle, penetrating the medial cortex of the pedicle. (B, D) Lateral view, superior vertebra, cannula number 1 (prevertebral leak): we can observe the tip of the trocar penetrating the anterior cortex of the vertebral body. Inferior vertebrae, cannula number 2 (epidural leak): the tip of the cannula stays behind the posterior cortex of the vertebral body. (A, B) We can observe the polymethylmethacrylate leaking antero laterally to the upper vertebra in a fusiform way (prevertebral leak). In the inferior vertebrae we can observe the leakage in the spinal canal in a laminar way (epidural leak).

18%, 200 mg/kg.

Later on we performed sample acquisition and histological analysis: the lumbar spine of each pig was harvested from a posterior approach, maintaining the anterior muscles.

The pedicle insertion points were identified. An extensive and careful laminectomy was performed to expose the cement and the dural sac (epidural leak), after the laminectomy we took a set of photographs to document the distribution of the polymethylmethacrylate in the epidural space. Later on we collected the dural sacs by cutting the roots near the foramina; the cement inside the psoas muscle (prevertebral leak) was found by manual palpation and then, was exposed. We took photographs of every specimen.

All the areas of the dural sac and of the muscle that had been in contact with the cement were marked with Chinese ink to make it easy to find those areas during microscopical observation. We use Chinese ink because it is widely used to mark tissues for histological analysis without inducing any change on dead tissues (Fig. 2).

All the samples were fixed in formaldehyde.

Every tissue marked with Chinese ink were then dehydrated with ethanol, fixed in wax, cut in 5 μ m slices with a microtome, and stained with H&E.

Under light microscopy, a senior pathologist inspected the samples, searching for the presence of inflammatory reaction, tissue necrosis, foreign body reaction, or any other histological alteration.

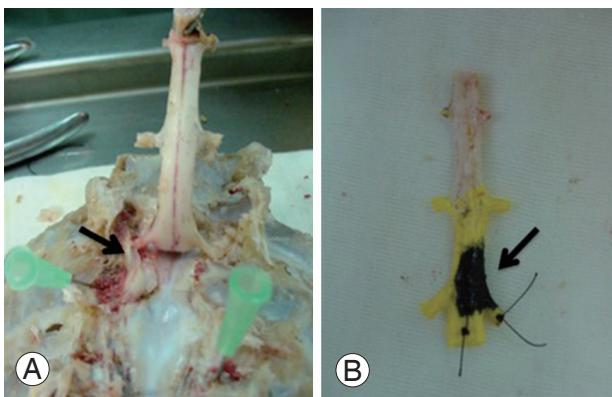


Fig. 2. (A) During removal of the spinal cord, we can see cement near the axilla of the root (arrow). (B) The area of the cord in contact with the cement was marked with ink (arrow).

Results

1. Clinical results

Pig number 5 presented a transient tachycardia during the surgery without any clinical consequence.

Pig number 6 died during the procedure after the cement injection. Necropsy did not reveal the cause of death. We especially looked for venous or pulmonary cement embolisms.

None of the pigs presented any neurological impairment after the procedure.

No superficial infections or healing problems were observed in the whole group of animals.

2. Macroscopic result

1) Epidural leak

The five vertebrae with epidural leak showed a laminar distribution of the polymerized cement that surrounded the dural sac and the root ipsilateral to the injection side, as a thin layer of cement (Fig. 3A). In one case the cement practically surrounded the cord a full 360° (Fig. 3B). Wide areas of the dural sacs that were in contact with the cement were successfully identified and were marked with Chinese ink. Those areas did not show any evidence of macroscopic changes.

2) Anterior prevertebral leak

The cement inside the muscles was distributed in a fusiform way by creating cavities. The inner layer of those cavities had a pseudo membrane aspect (Fig. 3C, D).

3. Histological results

1) Epidural leak

On dura mater we observed synovial metaplasia in three of five pigs, an inflammatory reaction (presence of lymphocytes, neutrophils, and eosinophils) in four of five, crystal deposits in four of five pigs and a giant cell reaction around crystal deposits in four of five pigs (Fig. 4).

The histological study of the spinal cord of the five animals, were completely normal (Fig. 4).

2) Anterior prevertebral leak (muscular findings)

In four out of the five pigs, inflammatory reaction was observed.

All the samples demonstrated crystal deposits, giant cell reaction, muscular atrophy, and synovial metaplasia (Fig. 5).

Discussion

We consider that the vertebroplasty model in pigs is very similar to the procedure that we normally used in our patients: the morphology of the vertebra and the size of the animal enabled us to perform the procedure using the same trocars that we used in humans. Moreover, the surgical setting is the same as the one we use for our patients.

Despite massive leakage of cement into the epidural space (Figs. 2, 3), none of the animals presented neu-

rological impairment after two weeks of observation. These findings are comparable to clinical series, in which

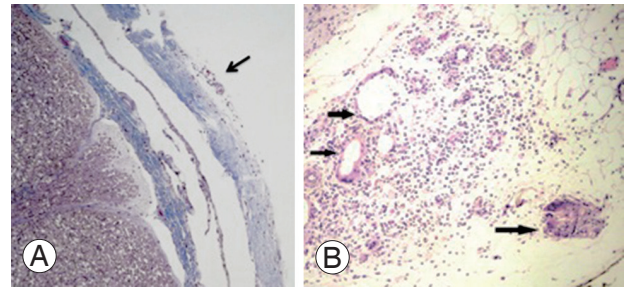


Fig. 4. Spinal cord and dura mater histology. (A) Normal spinal cord. The arrow indicates the dura mater with an inflammatory reaction (H&E, x2). (B) A close-up of the dura mater reveals a lot of lymphocytes; arrows indicate giant cell formations (H&E, x4).

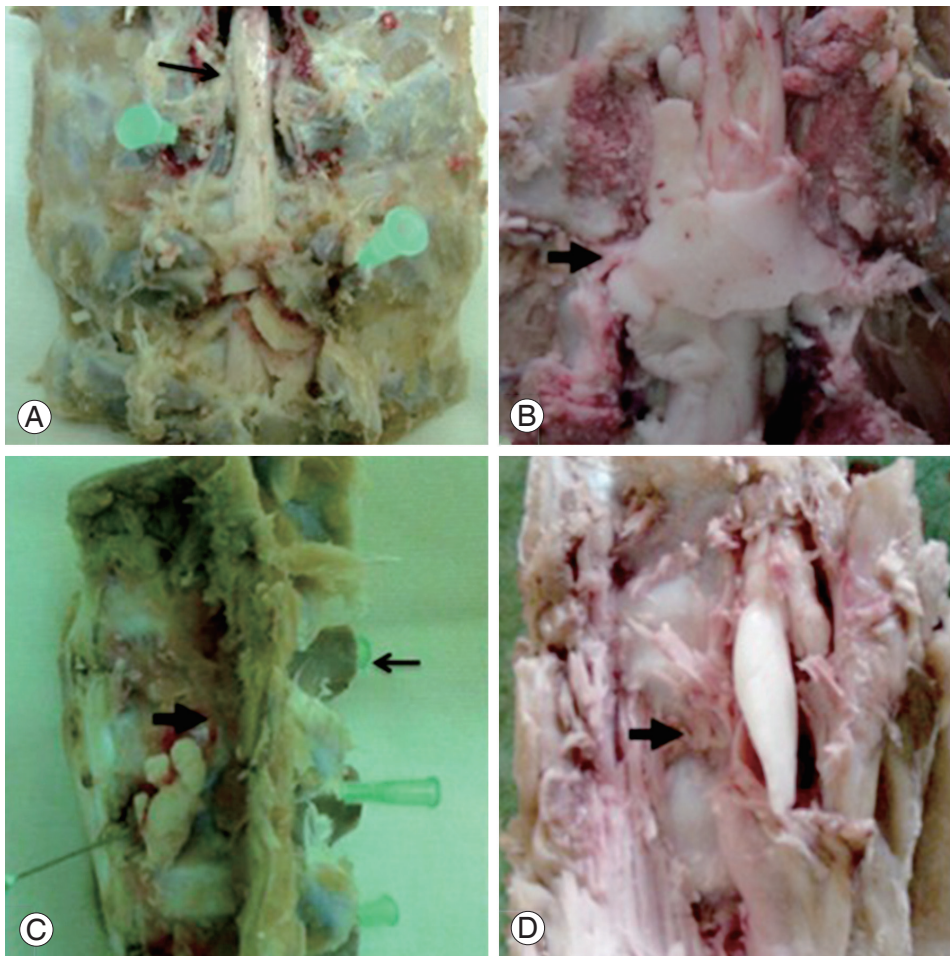


Fig. 3. Macroscopic findings. (A) Antero-posterior view of the spine. Pig number 1: post laminectomy, the needles show the trocars entering the pedicles, the arrow shows the cord. (B) Antero-posterior view of the spine. Pig number 3: post laminectomy, the arrow shows the spinal cord completely surrounded by cement. (C) Lateral view of the spine. Pig number 2: the thick arrow shows the cement in a prevertebral location; the thin arrow shows the spinous process. (D) Lateral view of the spine. Pig number 5: cement is observed in the prevertebral area inside the psoas (arrow).

radiological leaks are very common but neurological impairment that requires surgery are highly infrequent [11]. A possible explanation of this phenomenon is that the liquid cement is distributed in a laminar way around the dura mater, while is modeled by the pulsatile movements of the dura mater.

On the dura mater and the prevertebral muscles we observed an inflammatory reaction (presence of lymphocytes, neutrophils, and eosinophils), a foreign body reaction, and fibrosis. Those changes are probably related to the toxicity of polymethylmethacrylate, the presence of Barium crystals and the exothermal reaction of the cement during its polymerization [17-23].

We did not find necrosis on the dura mater, a probable explanation is that the polymerization temperatures may not reach the threshold of tissue necrosis in a live animal, probably because the epidural vessels and the cerebrospinal fluid are able to dissipate the heat and prevent the

exothermal damage [23,27].

We also did not observe any changes in the spinal cord, probably because the dura mater and cerebrospinal fluid are sufficiently able to isolate the neural structures from the inflammatory reaction and from the exothermic reaction of the curing cement.

On the muscular tissue we observed slight atrophy, fibrosis, and no necrosis. These changes differ from the changes described on bone, where the necrosis is more frequent. This could be because liquid cement may require less pressure to distribute itself, inside soft tissue than inside the bone; in fact, an *in-vitro* study in cadaveric vertebrae showed an increase of 13.6 folds on average over the base line of bone pressure during the injection of polymethylmethacrylate [28].

Synovial metaplasia was observed in both dura mater and muscles that had been in contact with the cement; the pulsatile rubbing between tissue and the cement may

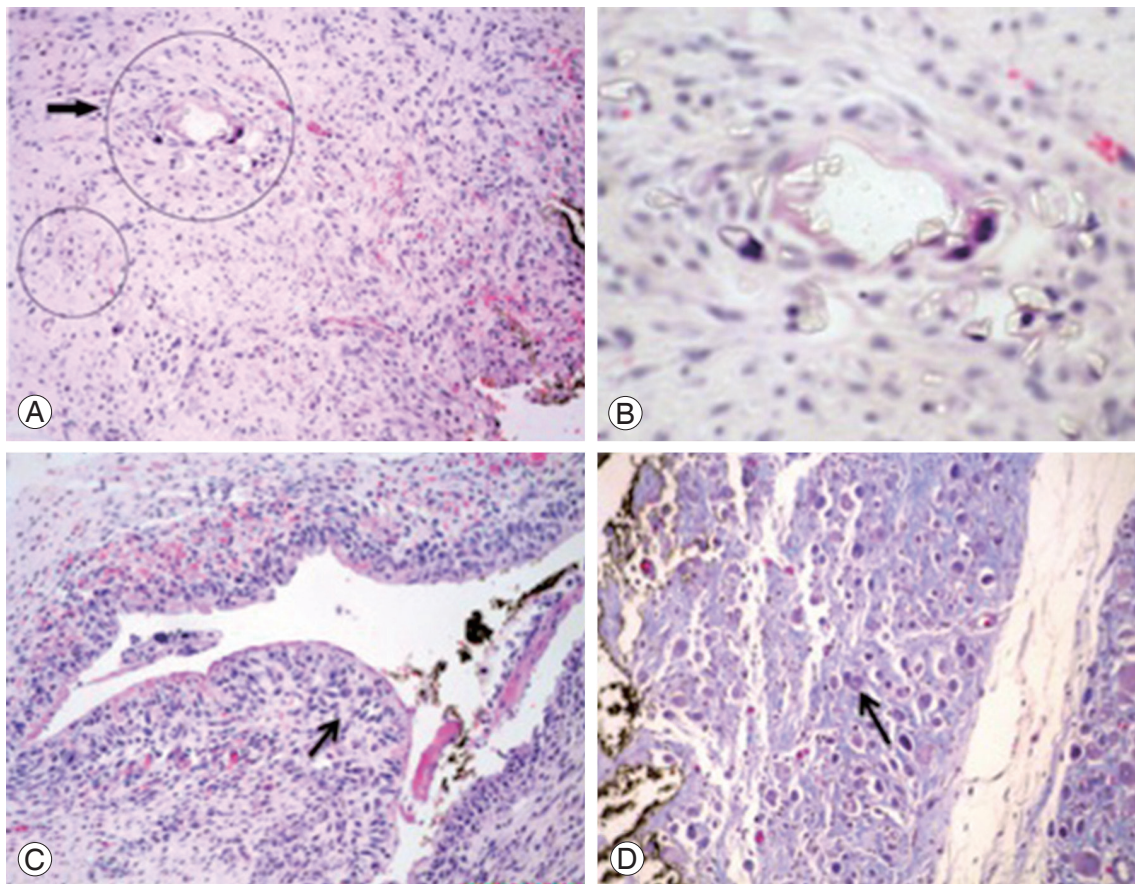


Fig. 5. Histology of the prevertebral muscle. (A) Muscle with inflammation, the arrow and circles indicate giant cell formation (H&E, $\times 4$). (B) Close-up, we can observe a giant cell and many crystals inside and around it (H&E, $\times 40$). (C) The arrow shows synovial metaplasia of the muscle near the ink mark (H&E, $\times 4$). (D) Muscle fibrosis and atrophy (arrow) (H&E, $\times 20$).

be responsible for this histological adaptation. As far as we know, synovial metaplasia is not previously described, probably because all the literature available focuses on changes in the bone.

Conclusions

The vertebroplasty model in pigs is very similar to the procedure in humans. This study shows that the way the cement distributes itself inside the epidural space and in perivertebral muscles does not induce a significant clinical alteration. The laminar way in which the cement is distributed in the epidural space may be what prevents a mechanical lesion of the spinal cord. We did not observe any changes in the spinal cord, probably because the dura mater and cerebrospinal fluid are sufficiently able to isolate the neural structure from the polymethylmethacrylate. We observed an inflammatory reaction, a foreign body reaction, fibrosis, and synovial metaplasia on the dura mater and on the prevertebral muscles.

Conflict of Interest

No potential conflict of interest relevant to this article was reported.

Acknowledgments

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References

- Galibert P, Deramond H, Rosat P, Le Gars D. Preliminary note on the treatment of vertebral angioma by percutaneous acrylic vertebroplasty. *Neurochirurgie* 1987;33:166-8.
- Barr JD, Barr MS, Lemley TJ, McCann RM. Percutaneous vertebroplasty for pain relief and spinal stabilization. *Spine (Phila Pa 1976)* 2000;25:923-8.
- Kallmes DF, Schweickert PA, Marx WF, Jensen ME. Vertebroplasty in the mid- and upper thoracic spine. *AJNR Am J Neuroradiol* 2002;23:1117-20.
- Anselmetti GC, Marcia S, Saba L, et al. Percutaneous vertebroplasty: multi-centric results from EVEREST experience in large cohort of patients. *Eur J Radiol* 2012;81:4083-6.
- Deramond H, Depriester C, Galibert P, Le Gars D. Percutaneous vertebroplasty with polymethylmethacrylate. Technique, indications, and results. *Radiol Clin North Am* 1998;36:533-46.
- Kallmes DF, Jensen ME. Percutaneous vertebroplasty. *Radiology* 2003;229:27-36.
- Chiras J, Depriester C, Weill A, Sola-Martinez MT, Deramond H. Percutaneous vertebral surgery: techniques and indications. *J Neuroradiol* 1997;24:45-59.
- Nussbaum DA, Gailloud P, Murphy K. A review of complications associated with vertebroplasty and kyphoplasty as reported to the Food and Drug Administration medical device related web site. *J Vasc Interv Radiol* 2004;15:1185-92.
- Padovani B, Kasriel O, Brunner P, Peretti-Viton P. Pulmonary embolism caused by acrylic cement: a rare complication of percutaneous vertebroplasty. *AJNR Am J Neuroradiol* 1999;20:375-7.
- Weill A, Chiras J, Simon JM, Rose M, Sola-Martinez T, Enkaoua E. Spinal metastases: indications for and results of percutaneous injection of acrylic surgical cement. *Radiology* 1996;199:241-7.
- Cotten A, Dewatre F, Cortet B, et al. Percutaneous vertebroplasty for osteolytic metastases and myeloma: effects of the percentage of lesion filling and the leakage of methyl methacrylate at clinical follow-up. *Radiology* 1996;200:525-30.
- Harrington KD. Major neurological complications following percutaneous vertebroplasty with polymethylmethacrylate: a case report. *J Bone Joint Surg Am* 2001;83:1070-3.
- Gaughen JR Jr, Jensen ME, Schweickert PA, Kaufmann TJ, Marx WF, Kallmes DF. Relevance of antecedent venography in percutaneous vertebroplasty for the treatment of osteoporotic compression fractures. *AJNR Am J Neuroradiol* 2002;23:594-600.
- Kaufmann TJ, Wald JT, Kallmes DF. A technique to circumvent subcutaneous cement tracts during percutaneous vertebroplasty. *AJNR Am J Neuroradiol* 2004;25:1595-6.
- Perez-Higuera A, Galovic LA. Percutaneous vertebroplasty: indications and technique. *España: Editorial Médica Panamericana*; 2003.
- Jung JY, Lee MH, Ahn JM. Leakage of polymethylmethacrylate in percutaneous vertebroplasty: comparison of osteoporotic vertebral compression

- fractures with and without an intravertebral vacuum cleft. *J Comput Assist Tomogr* 2006;30:501-6.
17. Verlaan JJ, Oner FC, Slootweg PJ, Verbout AJ, Dhert WJ. Histologic changes after vertebroplasty. *J Bone Joint Surg Am* 2004;86:1230-8.
 18. Togawa D, Kovacic JJ, Bauer TW, Reinhardt MK, Brodke DS, Lieberman IH. Radiographic and histologic findings of vertebral augmentation using polymethylmethacrylate in the primate spine: percutaneous vertebroplasty versus kyphoplasty. *Spine (Phila Pa 1976)* 2006;31:E4-10.
 19. Kobayashi N, Togawa D, Fujishiro T, et al. Histological and radiographic evaluation of polymethylmethacrylate with two different concentrations of barium sulfate in a sheep vertebroplasty model. *J Biomed Mater Res A* 2005;75:123-7.
 20. Huang KY, Yan JJ, Lin RM. Histopathologic findings of retrieved specimens of vertebroplasty with polymethylmethacrylate cement: case control study. *Spine (Phila Pa 1976)* 2005;30:E585-8.
 21. Togawa D, Bauer TW, Lieberman IH, Takikawa S. Histologic evaluation of human vertebral bodies after vertebral augmentation with polymethyl methacrylate. *Spine (Phila Pa 1976)* 2003;28:1521-7.
 22. Kim CW, Minocha J, Wahl CE, Garfin SR. Response of fractured osteoporotic bone to polymethylacrylate after vertebroplasty: case report. *Spine J* 2004;4:709-12.
 23. Urrutia J, Bono CM, Mery P, Rojas C. Early histologic changes following polymethylmethacrylate injection (vertebroplasty) in rabbit lumbar vertebrae. *Spine (Phila Pa 1976)* 2008;33:877-82.
 24. Flecknell P. Assessment and alleviation of post-operative pain [Internet]. Beltsville: Animal Welfare Information Center; 1998 [cited 2012 Sep 6]. Available from: <http://www.nal.usda.gov/awic/newsletters/v8n3/8n3fleck.htm>.
 25. Olfert E. Defining an acceptable endpoint in invasive experiments [Internet]. Beltsville: Animal Welfare Information Center; 1995 [cited 2012 Sep 6]. Available from: <http://www.nal.usda.gov/awic/newsletters/v6n1/6n1olfer.htm>.
 26. Tarlov IM. Spinal cord compression studies: III. Time limits for recovery after gradual compression in dogs. *AMA Arch Neurol Psychiatry* 1954;71:588-97.
 27. Verlaan JJ, Oner FC, Verbout AJ, Dhert WJ. Temperature elevation after vertebroplasty with polymethylmethacrylate in the goat spine. *J Biomed Mater Res B Appl Biomater* 2003;67:581-5.
 28. Gravius S, Kraska N, Maus U, Mumme T, Berdel P, Weisskopf M. Intravertebral pressure during vertebroplasty: an in-vitro study. *Z Orthop Unfall* 2009;147:43-7.